# Department of Biotechnology NPTEL/SWAYAM Courses for Honours Degree in Biotechnology

| SI.<br>No | Code     | Course Name  | Duration | Credits | Link   |
|-----------|----------|--|----------|---------|--|
| 1         | H20BT001 | Solid and hazardous Waste<br>Management  | 15 Weeks | 4       | https://swayam.gov.in/nd2_cec20<br>_ge34/preview           |
| 2         | H20BT002 | Dairy and Food process and<br>products technology  | 12 Weeks | 3       | https://swayam.gov.in/nd1_noc20<br>_ag02/preview           |
| 3         | H20BT003 | Thermal Processing of foods  | 12 Weeks | 3       | https://swayam.gov.in/nd1_noc20<br>_ag07/preview_          |
| 4         | H20BT004 | Food Safety and Quality<br>Control   | 8 Weeks  | 2       | https://swayam.gov.in/nd2_cec20<br>_ag06/preview           |
| 5         | H20BT005 | Organic farming for<br>Sustainable Agricultural<br>Production  | 8 Weeks  | 2       | https://swayam.gov.in/nd1_noc20<br>_ag05/preview           |
| 6         | H20BT006 | Introduction To<br>Mechanobiology  | 8 Weeks  | 2       | https://swayam.gov.in/nd1_noc20<br>_bt27/preview           |
| 7         | H20BT007 | Biomedical Nanotechnology  | 4 Weeks  | 1       | https://swayam.gov.in/nd1_noc20<br>_bt29/preview           |
| 8         | H20BT008 | Biomicrofluidics   | 4 Weeks  | 1       | https://swayam.gov.in/nd1_noc20<br>_bt22/preview           |
| 9         | H20BT009 | Fundamentals of Micro And<br>Nanofabrication   | 12 Weeks | 3       | https://swayam.gov.in/nd1_noc20<br>_bt37/preview_          |
| 10        | H20BT010 | Industrial pharmacy I  | 12 Weeks | 3       | https://swayam.gov.in/nd2_cec20<br>_lb05/preview           |
| 11        | H20BT011 | Mechanical Unit Operations   | 12 Weeks | 3       | https://swayam.gov.in/nd1_noc20<br>ch27/preview_           |
| 12        | H20BT012 | Nanotechnology In<br>Agriculture   | 8 Weeks  | 2       | https://swayam.gov.in/nd1_noc20<br>_bt41/preview           |
| 13        | H20BT013 | Introduction to Biomaterials<br>(being offered as professional<br>elective in regular<br>curriculum) | 8 Weeks  | 2       | https://nptel.ac.in/content/syllabus<br>_pdf/113104009.pdf |
| 14        | H20BT014 | Wildlife Conservation  | 8 Weeks  | 2       | https://swayam.gov.in/nd1_noc20<br>_bt39/preview           |
| 15        | H20BT015 | Computational systems<br>Biology   | 12 Weeks | 3       | https://nptel.ac.in/content/syllabus<br>_pdf/102106068.pdf |
| 16        | H20BT016 | Drug Delivery: Principles  | 12 Weeks | 3       | https://swayam.gov.in/nd1_noc20                            |

|    |          | And Engineering  |          |   | _bt24/preview  |
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| 17 | H20BT017 | Introduction To<br>Proteogenomics  | 12 Weeks | 3 | https://swayam.gov.in/nd1_noc20<br>_bt19/preview         |
| 18 | H20BT018 | Patent Law for Engineers and<br>Scientists   | 12 Weeks | 3 | https://swayam.gov.in/nd1_noc20<br>_hs55/preview         |
| 19 | H20BT019 | Computer Aided Drug Design   | 8 Weeks  | 2 | https://swayam.gov.in/nd1_noc20<br>_bt23/preview         |
| 20 | H20BT020 | Introduction To Proteomics<br>(being offered as professional<br>elective in regular<br>curriculum)     | 8 Weeks  | 2 | https://swayam.gov.in/nd1_noc20<br>_bt20/preview         |
| 21 | H20BT021 | Introduction to cell culture<br>Technology   | 8 Weeks  | 2 | https://nptel.ac.in/courses/102/104<br>/102104059/       |
| 22 | H21BT022 | Maternal Infant Young Child<br>Nutrition   | 12 Weeks | 3 | https://onlinecourses.nptel.ac.in/n<br>oc22_bt01/preview |
| 23 | H21BT023 | Optical Spectroscopy and<br>Microscopy: Fundamentals of<br>optical measurements and<br>instrumentation | 12 Weeks | 3 | https://onlinecourses.nptel.ac.in/n<br>oc22_bt05/preview |
| 24 | H21BT024 | Human Molecular Genetics   | 4 Weeks  | 1 | https://onlinecourses.nptel.ac.in/n<br>oc22_bt07/preview |
| 25 | H21BT025 | Demystifying The Brain   | 4 Weeks  | 1 | https://onlinecourses.nptel.ac.in/n<br>oc22_bt14/preview |
| 26 | H21BT026 | Medical Biomaterials   | 8 Weeks  | 2 | https://onlinecourses.nptel.ac.in/n<br>oc22_bt15/preview |
| 27 | H21BT027 | Forests And Their<br>Management  | 12 Weeks | 3 | https://onlinecourses.nptel.ac.in/n<br>oc22_bt24/preview |
| 28 | H21BT028 | Bioelectrochemistry  | 4 Weeks  | 1 | https://onlinecourses.nptel.ac.in/n<br>oc22_bt26/preview |
| 29 | H21BT029 | Plant Developmental Biology  | 4 Weeks  | 1 | https://onlinecourses.nptel.ac.in/n<br>oc22_bt27/preview |
| 30 | H21BT030 | Conservation Economics   | 12 Weeks | 3 | https://onlinecourses.nptel.ac.in/n<br>oc22_bt31/preview |
| 31 | H21BT031 | Soft Nano Technology   | 8 Weeks  | 2 | https://onlinecourses.nptel.ac.in/n<br>oc22_ch11/preview |
| 32 | H21BT032 | Understanding Design   | 4 Weeks  | 1 | https://onlinecourses.nptel.ac.in/n<br>oc22_de01/preview |

| 33 | H21BT033 | Design, Technology and<br>Innovation   | 8 Weeks  | 2 | https://onlinecourses.nptel.ac.in/n<br>oc22_de05/preview  |
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| 34 | H21BT034 | Emotional Intelligence   | 8 Weeks  | 2 | https://onlinecourses.nptel.ac.in/n<br>oc22_hs11/preview  |
| 35 | H21BT035 | Exploring Survey Data on<br>Health Care  | 8 Weeks  | 2 | https://onlinecourses.nptel.ac.in/n<br>oc22_hs40/preview  |
| 36 | H21BT036 | Material and Energy Balances   | 12 Weeks | 3 | https://onlinecourses.nptel.ac.in/n<br>oc22_bt04/preview  |
| 37 | H21BT037 | Bioreactor Design and<br>Analysis  | 8 Weeks  | 2 | https://onlinecourses.nptel.ac.in/n<br>oc22_bt19/preview  |
| 38 | H21BT038 | Waste to Energy Conversion   | 8 Weeks  | 2 | https://onlinecourses.nptel.ac.in/n<br>oc22_ch05/preview  |
| 39 | H21BT039 | Physico-chemical processes<br>for wastewater treatment   | 12 Weeks | 3 | https://onlinecourses.nptel.ac.in/n<br>oc22_ch25/preview  |
| 40 | H21BT040 | Renewable Energy<br>Engineering: Solar, Wind and<br>Biomass Energy Systems                               | 12 Weeks | 3 | https://onlinecourses.nptel.ac.in/n<br>oc22_ch27/preview  |
| 41 | H21BT041 | Biomass Conversion and<br>Biorefinery  | 12 Weeks | 3 | https://onlinecourses.nptel.ac.in/n<br>oc22_ch28/preview  |
| 42 | H21BT042 | Environmental Quality<br>Monitoring & Analysis   | 12 Weeks | 3 | https://onlinecourses.nptel.ac.in/n<br>oc22_ch33/preview  |
| 43 | H21BT043 | Bio photonics  | 12 Weeks | 3 | https://onlinecourses.nptel.ac.in/n<br>oc22_ee59/preview  |
| 44 | H21BT044 | Introduction to<br>Environmental Engineering<br>and Science - Fundamental<br>and Sustainability Concepts | 12 Weeks | 3 | https://onlinecourses.nptel.ac.in/n<br>oc22_ge06/preview  |
| 45 | H21BT045 | Interactomics: Basics &<br>Applications  | 12 Weeks | 3 | https://onlinecourses.nptel.ac.in/n<br>oc22_bt11/preview  |
| 46 | H21BT046 | Data Analysis for Biologists   | 8 Weeks  | 2 | https://onlinecourses.nptel.ac.in/n<br>oc22_bt20/preview  |
| 47 | H21BT047 | Structural Biology   | 12 Weeks | 3 | https://onlinecourses.nptel.ac.in/n<br>oc22_bt28/preview  |
| 48 | H21BT048 | Biointerface Engineering   | 8 Weeks  | 2 | https://onlinecourses.nptel.ac.in/n<br>oc22_bt21/preview  |
| 49 | H21BT049 | Introduction to Dynamical<br>Models in Biology   | 4 Weeks  | 1 | https://onlinecourses.nptel.ac.in/n<br>oc 21_bt42/preview |

| 50 | H21BT050 | Conservation Geography   | 12 Weeks | 2 | https://onlinecourses.nptel.ac.in/n<br>oc21_bt44/preview   |
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| 51 | H21BT051 | Wildlife Ecology   | 12 Weeks | 3 | https://onlinecourses.nptel.ac.in/n<br>oc 21_bt45/preview  |
| 52 | H21BT052 | Organic Chemistry in Biology<br>and Drug Development   | 12 Weeks | 3 | https://onlinecourses.nptel.ac.in/n<br>oc 21_cy30/preview  |
| 53 | H21BT053 | Legal and Regulatory Issues<br>in Biotechnology  | 4 Weeks  | 1 | https://onlinecourses.nptel.ac.in/n<br>oc 21_lw04/preview  |
| 54 | H21BT054 | Nanomaterials and their<br>Properties  | 12 Weeks | 3 | https://onlinecourses.nptel.ac.in/n<br>oc21_mm38/preview   |
| 55 | H21BT055 | Ecology and Environment  | 8 Weeks  | 2 | https://onlinecourses.nptel.ac.in/n<br>oc 21_ge16/preview  |
| 56 | H21BT056 | Bioreactors  | 4 Weeks  | 1 | https://onlinecourses.nptel.ac.in/n<br>oc21_bt28/preview   |
| 57 | H21BT057 | Transport Phenomena in<br>Biological Systems   | 12 Weeks | 3 | https://onlinecourses.nptel.ac.in/n<br>oc 21_bt40/preview  |
| 58 | H21BT058 | Technologies For Clean And<br>Renewable Energy<br>Production   | 8 Weeks  | 2 | https://onlinecourses.nptel.ac.in/n<br>oc 21_ch42/preview  |
| 59 | H21BT059 | Principles and Practices of<br>Process Equipment and Plant<br>Design   | 12 Weeks | 3 | https://onlinecourses.nptel.ac.in/n<br>oc 21_ch52/preview  |
| 60 | H21BT060 | Introduction to Biomedical<br>Imaging Systems  | 12 Weeks | 3 | https://onlinecourses.nptel.ac.in/n<br>oc 21_bt50/preview  |
| 61 | H21BT061 | Biomechanics of Joints and<br>Orthopaedic Implants   | 8 Weeks  | 2 | https://onlinecourses.nptel.ac.in/n<br>oc 21_me130/preview |
| 62 | H21BT062 | Neuroscience of Human<br>Movements   | 12 Weeks | 3 | https://onlinecourses.nptel.ac.in/n<br>oc21_ge17/preview   |
| 63 | H21BT063 | Current regulatory<br>requirements for conducting<br>clinical trials in India for<br>investigational new<br>drugs/new drug (Version 3.0) | 8 Weeks  | 2 | https://onlinecourses.nptel.ac.in/n<br>oc 21_ge25/preview  |
| 64 | H21BT064 | Functional Genomics  | 4 Weeks  | 1 | https://onlinecourses.nptel.ac.in/n<br>oc21_bt39/preview   |
| 65 | H21BT065 | Aspects of Biochemical<br>Engineering  | 12 Weeks | 3 | https://onlinecourses.nptel.ac.in/<br>noc21_bt17/preview   |

| 66 | H21BT066 | RNA Biology  | 12 Weeks | 3 | https://nptel.ac.in/courses/102106<br>097                |
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| 67 | H21BT067 | Biomechanics   | 12 Weeks | 3 | https://nptel.ac.in/courses/102106<br>098                |
| 68 | H21BT068 | Enzyme Sciences and<br>Technology  | 12 Weeks | 3 | https://nptel.ac.in/courses/102103 097                   |
| 69 | H21BT069 | Metabolic Engineering  | 8 Weeks  | 2 | https://nptel.ac.in/courses/102105<br>086                |
| 70 | H21BT070 | Introduction to Professional<br>Scientific Communication   | 4 Weeks  | 1 | https://nptel.ac.in/courses/102104<br>061                |
| 71 | H21BT071 | Bioengineering: An Interface<br>with Biology and Medicine  | 8 Weeks  | 2 | https://nptel.ac.in/courses/102101<br>068                |
| 72 | H24BT072 | Environmental Biotechnology<br>(being offered as Elective in<br>our R20 curriculum)              | 12 Weeks | 3 | https://onlinecourses.nptel.ac.in/n<br>oc23_bt60/preview |
| 73 | H24BT073 | Industrial Biotechnology<br>(being offered as Elective in<br>our R20 curriculum)                 | 12 Weeks | 3 | https://onlinecourses.nptel.ac.in/n<br>oc23_bt61/preview |
| 74 | H24BT074 | Introduction To Biostatistics  | 8 Weeks  | 2 | https://onlinecourses.nptel.ac.in/n<br>oc23_bt58/preview |
| 75 | H24BT075 | Medical Image Analysis   | 12 Weeks | 3 | https://onlinecourses.nptel.ac.in/n<br>oc23_bt40/preview |
| 76 | H24BT076 | Introduction To<br>Developmental Biology<br>(being offered as Elective in<br>our R20 curriculum) | 12 Weeks | 3 | https://onlinecourses.nptel.ac.in/n<br>oc23_bt43/preview |
| 77 | H24BT077 | Tissue Engineering (being<br>offered as Elective in our R20<br>curriculum)                       | 8 Weeks  | 2 | https://onlinecourses.nptel.ac.in/n<br>oc23_bt46/preview |
| 78 | H24BT078 | Organ Printing   | 8 Weeks  | 2 | https://onlinecourses.nptel.ac.in/n<br>oc23_bt49/preview |
| 79 | H24BT079 | Genome Editing And<br>Engineering  | 12 Weeks | 3 | https://onlinecourses.nptel.ac.in/n<br>oc23_bt51/preview |
| 80 | H24BT080 | Next Generation Sequencing<br>Technologies : Data Analysis<br>And Applications                   | 12 Weeks | 3 | https://onlinecourses.nptel.ac.in/n<br>oc23_bt34/preview |
| 81 | H24BT081 | Computational Neuroscience   | 12 Weeks | 3 | https://onlinecourses.nptel.ac.in/n<br>oc23_bt64/preview |

| 82 | H24BT082 | Neurobiology  | 4 weeks  | 1 | https://onlinecourses.nptel.ac.in/n<br>oc23_bt65/preview |
|----|----------|---|----------|---|--|
| 83 | H24BT083 | Neural Science for Engineers  | 12 Weeks | 3 | https://onlinecourses.nptel.ac.in/n<br>oc24_ee01/preview |
| 84 | H24BT084 | Classics in Neuroscience  | 8 Weeks  | 2 | https://onlinecourses.nptel.ac.in/n<br>oc24_bt38/preview |
| 85 | H24BT085 | Human Behaviour   | 8 Weeks  | 2 | https://onlinecourses.nptel.ac.in/n<br>oc24_hs35/preview |
| 86 | H24BT086 | Microsensors, Implantable<br>Devices and Rodent Surgeries<br>for Biomedical Applications                | 12 Weeks | 3 | https://onlinecourses.nptel.ac.in/n<br>oc24_bt37/preview |
| 87 | H24BT087 | Education for Sustainable<br>Development  | 12 Weeks | 3 | https://onlinecourses.nptel.ac.in/n<br>oc24_hs04/preview |
| 88 | H24BT088 | Advances in Omics   | 8 Weeks  | 2 | https://onlinecourses.nptel.ac.in/n<br>oc24_bt42/preview |
| 89 | H24BT089 | Statistics for Biomedical<br>Engineers  | 12 Weeks | 3 | https://onlinecourses.nptel.ac.in/n<br>oc24_bt70/preview |
| 90 | H24BT090 | Design for Biosecurity  | 12 Weeks | 3 | https://onlinecourses.nptel.ac.in/n<br>oc24_bt77/preview |
| 91 | H24BT091 | Advanced Fluorescence<br>Microscopy and Image<br>Processing   | 12 Weeks | 3 | https://onlinecourses.nptel.ac.in/n<br>oc25_bt01/preview |
| 92 | H24BT092 | Computational Genomics  | 12 Weeks | 3 | https://onlinecourses.nptel.ac.in/n<br>oc25_bt13/preview |
| 93 | H24BT093 | Microbial Biotechnology   | 12 Weeks | 3 | https://onlinecourses.nptel.ac.in/n<br>oc25_bt33/preview |
| 94 | H24BT094 | Optical Spectroscopy and<br>Microscopy : Fundamentals<br>of Optical Measurements and<br>Instrumentation | 12 Weeks | 3 | https://onlinecourses.nptel.ac.in/n<br>oc25_bt37/preview |
| 95 | H24BT095 | Biological Data Analysis and<br>Visualization with R  | 8 Weeks  | 2 | https://onlinecourses.nptel.ac.in/n<br>oc25_bt43/preview |
| 96 | H24BT096 | Experimental<br>Nanobiotechnology   | 4 Weeks  | 1 | https://onlinecourses.nptel.ac.in/n<br>oc25_bt44/preview |
| 97 | H24BT097 | Algorithms in Computational<br>Biology and Sequence<br>Analysis   | 12 Weeks | 3 | https://nptel.ac.in/courses/106108<br>571                |

| 98  | H24BT098 | Biophotonics  | 12 Weeks | 3 | https://onlinecourses.nptel.ac.in/n<br>oc24_ge11/preview |
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| 99  | H24BT099 | Comprehensive Molecular<br>Diagnostics and Advanced<br>Gene Expression Analysis | 12 Weeks | 3 | https://onlinecourses.nptel.ac.in/n<br>oc24_ge36/preview |
| 100 | H24BT100 | Cellular Biophysics: A<br>Framework For Quantitative<br>Biology                 | 8 weeks  | 2 | https://onlinecourses.nptel.ac.in/n<br>oc22_bt32/preview |

## Solid and hazardous Waste Management

## Duration: 15 weeks Credits: 4

## **Course layout**

## Week 01

01. Waste- Types and classification

02. Waste sources and generation rates

03. Traditional methods of waste collection and disposal

## Week 02

04. Factors influencing waste generation and health hazards05. Waste composition06. Waste collection - I

### Week 03

07. Waste collection - II 08. Characterization of wastes

### Week 04

09. Waste processing : Size and volume reduction

10. Waste minimization, waste hierarchy and waste audit

## Week 05

11. Recycling of solid wastes

12. Hazardous waste- Definition, sources, classification, collection and segregation

13. Hazardous waste characterization, treatment and disposal

### Week 06

14. Radioactive waste-I

15. Radioactive waste-II

### Week 07

16. e waste17. Plastic waste

Week 08 18. Biomedical waste- I Biomedical waste-II
 Biomedical waste management rules, 2016

## Week 09

21. Composting

22. Vermicomposting

23. Biogas production from solid waste

# Week 10

24. Thermal treatment of solid waste - Incineration

25. Thermal treatment of solid waste - Pyrolysis and gasification

# Week 11

26. Solid waste disposal - Sanitary landfilling- I

27. Solid waste disposal - Sanitary landfilling- II

## Week 12

- 28. Landfill leachate and gas management
- 29. Landfill bioreactors
- 30. Fly ash- Generation and management
- 31. Fly ash management
- 32. Noti/Solid waste management rules, 2016
- 33. Hazardous and other waste (management and transboundary movement) amendment rules, 2016
- 34. Plastic waste management rules, 2016

# Week 14

35. e waste management rules, 2016

- 36. Basel Convention on the control of transboundary movement of hazardous waste and their disposal
- 37. Mechanical biological treatment of solid wastes

## Week 15

- 38. Solid waste management in rural areas
- 39. Swachh Bharat Abhiyan
- 40. Recent advances in solid waste management

## Dairy and Food process and products technology

Duration: 12 weeks Credits: 3

## **Course layout**

Week 1 : Basic principles and methods of food processing and preservation. EmergingTechnologies in food processing. Food additives and preservatives.

Week 2 : Food lawsand standards. Effect of processing on acceptability and nutritive value offood.

Week 3 : Physico-chemical properties and structure of milk and milk constituents.

Week 4 : Chemical and microbial spoilage of milk and milk products; Fluid milkProcessing, packaging and distribution.

Week 5 : Common dairy processes – creamseparation (standardization), pasteurization, sterilization and Homogenization.

Week 6 : Process technology for manufacture of evaporated milk, condensedmilk, dried milk, malted milk, infant and baby foods, ice-cream, cheese, butter, fermented milk and indigenous dairy products.

Week 7 : Methods and procedures for sampling and testing of milk and milk products. Laws and standards for milk and milk products.

Week 8 : Technological processes for industrially manufactured foods of commercial importance, from plant and animal origin.

Week 9 : Cereals, vegetables, fruits, meats, poultry and egg products; Bakery, pasta and confectionary products, ready to eat foods, fermented foods, alcoholic and non- alcoholic Beverages, tea, coffee and cocoa, fabricated foods.

Week 10 : Packaging materials; Characteristics, properties and their design. Packaging requirement forDifferent processed and unprocessed foods.

Week 11 : Working Principles of various typeof fillers : form-fill- seal machine.

**Week 12** : Gas packaging and modified atmospherePackage design. Shelf life prediction of foods in packages. Quality control inFood packaging. Product safety and packaging regulations.

## **Books and references**

Nil

# **Thermal Processing of foods**

Duration: 12 weeks Credits: 3

### **Course layout**

Week 1 :Food microbiology: microbial growth and concerns in various foods, Blanching, Pasteurization, Ultrapasteurization, Hot fill and UHT

Week 2 : Thermal processing equipment, Milk pasteurization, Canning operations

Week 3 : Temperature distribution and heat penetration, Kinetics of reactions, F value and process requirements

Week 4 : Quality considerations and process optimization, Shelf life studies, Validation of heat processes

Week 5 : Fundamentals of aseptic processing, Aseptic equipment design, Aseptic process design

Week 6: Microwave and radio frequency heating, Ohmic heating, Overview of non-thermal processing technologies

Week 7:Advanced separation processes, High pressure, dialysis, ultrafiltration and reverse osmosis, Nanofiltration, electro dialysis and membrane separation

Week 8 :Various types of heat exchangers for food process engineering, Various types of driers for food process engineering

Week 9: Importance and applications of extrusion technology in food processing, Changes of properties and functional components of extruded foods

Week 10: Food biosensors, Types of functional foods: Probiotics and nutraceuticals

Week 11: Packaging considerations: Barrier and mechanical properties of different food packaging materials, Biocomposite/bionanocomposite materials for food packaging applications

Week 12: Sanitary components and requirements, Regulatory considerations

## **Books and references**

- 1. Chan, E. C. S., Pelczar, M. J., Jr. Krieg N. R. 2010. Microbiology. 5th edition. Tata McGraw Hill, Delhi.
- 2. Banwart, G.J. 1989. Basic Food Microbiology. 2nd edition. Chapman & Hall, New York.
- 3. Meyer, L.H. 2004. Food Chemistry. 1st edition. CBS Publishers & Distributors, Delhi.
- 4. McCabe, W. L., Smith J., Harriot P. 2014. Unit Operations of Chemical Engineering. 7th edition. McGraw Hill, International.
- 5. Nelson, P.E. (Editor). 2010. Principles of Aseptic Processing and Packaging. 3rd edition. Purdue university press.
- 6. Sun, D. (Editor). 2005. Emerging Technologies for Food Processing. Elsevier Academic Press.
- 7. Metaxas, A.C., Meridith, R.J. 1993. Industrial Microwave Heating. Peter Pergrinus Ltd., London.
- 8. Soroka, W. 1999. Fundamentals of Packaging Technology. Institute of Packaging Professionals.
- 9. Curtis, P.A. 2005. Guide to food laws regulations. Blackwell Publishing.

# **Food Safety and Quality Control**

### **Duration: 12 weeks** Credits: 3

## **Course layout**

#### Week 1

Assignments (No. & Type)\*

- 1. Food supplementation, substitution, fortification and enrichment
- 2. Importance of the costing of the Product
- Quality control and Internal control in Food Industry 3.
- 4. Sensory evaluation of food samples and container evolution

#### Week 2

Assignments (No. & Type)\*

Assignments (No. & Type)\*

- 1. Food Analysis
- 2. Waste control and sanitation Government regulations for quality standards
- 3. Food Standards
- 4. Primary Sources of Microorganisms in Food

#### Week 3

- 1. Fundamentals of control of microorganisms in foods
- 2. Food borne infections
- 3. Transmission of food borne infections
- 4. Microbes used in food biotechnology

### Week 4

Assignments (No. & Type)\*

- 1. Importance of sanitation and hygiene in food
- 2. Bacteriological analysis of food
- 3. Bacteriological analysis of water and milk
- 4. Morphological identification of molds and yeasts in foods

#### Week 5

Assignments (No. & Type)\*

- 1. Sampling to study the source of transmission of microorganisms in foods
- 2. Laws governing food service establishments
- 3. Laws concerning hygiene and safety
- 4. Hygiene, sanitation and safety of quantity food production

#### Week 6

Assignments (No. & Type)\*

- 1. Determinants of Health
- 2. Role of agricultural production in food availability and consumption
- 3. Post harvest handling, Marketing and Distribution of Foods
- 4. Food Adulteration

#### Week 7

- 1. Water pollution and its effects on food quality
- 2. Industrial effluents and their impact on food quality, Pesticide residues in foods
- 3. Food cost control in catering business
- 4. Recording and Reporting control charts Production control

#### Week 8

Assignments (No. & Type)\*

- 1. Menu planning, purchasing and storage of food for quantity food production
- 2. Acquisition of resources and organization

### **Books and references**

- Bhatti,S1995.Vame, Fruit and vegetable processing. CBS Publishers, Distributors, NewDelhi.
- Coles, R., McDowell, D., Kirwan, M.J. 2003. Food Packaging Technology. Blackwell Publishing Co.

◆ Dauthy, M. E. 1995. Fruits and Vegetables Processing- FAO Bulletin 119. International Book Distributing Co., Lucknow.

Devendra, K. B. and Priyanka, T. 2006. An Introduction to Food Science and technology and Quality Management.
 Kalyani Publishers 81-272-2521-5.

♣ FAO - Training Manual No.17/2. 2007. Prevention of post-harvest food losses: Fruits, Vegetables and Root crops. Daya Publishing House, Delhi.

- Fellows, P. J. 1998. Food Processing Technology principles and Practices. Ellis Horwood.
- Girdhari Lal, G.S. Siddappa and G.L. Tandon. 1959. Preservation of Fruits and Vegetables. ICAR, New Delhi.
- Girdharilal, Siddappa, G. S. and Tandon, G. L. 1998. Preservation of fruits and vegetables. ICAR, NewDelhi.
- Gordon L. Robertson. 2014. Food Packaging: Principles and Practice, 3rd Ed. CRC Press, Boca Raton, FL, USA.
- Gosby, N.T. 2001. Food Packaging Materials. Applied Science Publication

James G. Brennan. 2006. Food Processing Handbook. Wiley-VCH Verlag GmbH & Co. KGaA, Weinheim, Germany.

- John, P.J. 2008. A Handbook on Food Packaging Narendra Publishing House,
- Kalia, M. and Sood, S. 2010. Food Preservation and Processing. Revised Edition, Kalyani Publishers, New Delhi.
- KaysandStanely,J.1998. Postharvest physiology of perishable plant products. CBS Publishers, Distributors, New Delhi
- Mahadevia, M., Gowramma, R.V. 2007. Food Packaging Materials. Tata McGraw Hill
- Manoranjan, K. and Sangita, S. 1996. Food Preservation and Processing. Kalyani Publishers 978-81-272-4262-6.
- Monoranjam, K. and Sangita, S. 2008. Food Preservation and Processing. Kalyani Publishers 978-81-272-4262-6.
- P. Fellows. 2000. Food Processing Technology: Principles and Practice, 2nd Ed. CRC Press, Boca Raton, FL, USA.

- Robertson, G. L. 2001. Food Packaging and Shelf life: A Practical Guide. Narendra Publishing House.
- Robertson, G. L. 2005. Food Packaging: Principles and Practice. Second Edition. Taylor and Francis Pub.

S. Rajarathnam and R.S. Ramteke, 2011.Advances in preservation and processing technologies of fruits and vegetables.

Salunkhe, D.K., Bolin, H. R. and Reddy, N. R. 1991. Storage, Processing and Nutritional Quality of Fruits and Vegetables. 2nd Edition. Vol. II. CRC Press 0849356245

- Siddappaa, G. S., Girdhari Lal and Tandon, G.L. 1998. Preservation of Fruits and Vegetables. ICAR, New Delhi
- Sivasankar, B. 2002. Food Processing and Preservation. PHI Learning Pvt. Ltd. Delhi
- Srilakshmi. 2010. Food Science. New age International 978-81-224-2724-0.

Srivastava, R. P. & Sanjeev Kumar. 2002. Fruits and vegetable Preservation – Principles and Practice. International Book Distributing Co., Lucknow.

Swaminathan, M. 1988. Hand book of Food Science & Experimental Foods. Bappco publishers, Bangalore

U.D. Chavan and J.V. Patil. 2013. Industrial Processing of fruits and vegetables. Astral International Pvt Ltd. New Delhi.

Vijay, K. 2001. Text Book of Food Sciences and Technology. ICAR, New Delhi.

## **Organic farming for Sustainable Agricultural Production**

Duration: 8 weeks Credits: 2

## **Course layout**

- Week 1 : Organic Farming: Concepts and principles of organic farming
- Week 2 : Key indicators of sustainable agriculture, organic farming and climate change
- Week 3 : Input management; compost production, vermicomposting, Compost quality, Compost utilization and marketing
- Week 4 : Organic crop management: field crops, horticulture and plantation crops
- Week 5 : Plant protection measures, biopesticides, natural predators, cultural practice
- Week 6 : Rotation design for organic system, Transition to organic agriculture, farming system
- Week 7 : Quality analysis of organic foods, Antioxidants and their natural source, organic food and human health
- Week 8 : Standards of organic food and marketing

Books and references

Nil

### Introduction to Mechanobiology

### Duration: 8 weeks Credits: 2

## **Course layout**

### Week 1

Lecture 1: Need to study Mechanobiology Lecture 2: Cell as a Tent, individual components Lecture 3: Cell-ECM crosstalk Lecture 4: ECM proteins: Collagen Lecture 5: Measuring properties of collagen networks

#### Week 2

- Lecture 6: Properties of collagen networks
- Lecture 7: Rheology
- Lecture 8: Rheology of biopolymer networks
- Lecture 9: Atomic Force Microscopy (AFM)
- Lecture 10: Design of protein constructs for AFM

#### Week 3

- Lecture 11: Protein unfolding using AFM
- Lecture 12: Protein unfolding using AFM
- Lecture 13: Focal adhesions: focal adhesion proteins
- Lecture 14: Focal adhesion organization
- Lecture 15: Focal adhesions: role of forces

### Week 4

- Lecture 16: Cytoskeleton: Actin
- Lecture 17: Force-velocity relationships of actin networks
- Lecture 18: Mesenchymal cell migration
- Lecture 19: Actin dynamics during mesenchymal migration
- Lecture 20: Actin dynamics during mesenchymal migration

#### Week 5

- Lecture 21: Adhesion Independent Migration
- Lecture 22: Adhesion Independent & Collective Cell Migration
- Lecture 23: Collective Cell Migration
- Lecture 24: Mechanobiology of Stem Cell Fate I
- Lecture 25: Mechanobiology of Stem Cell Fate II

#### Week 6

- Lecture 26: Mechanobiology of Stem Cell Fate III
- Lecture 27: Mechanobiology of Diseases: Cancer I
- Lecture 28: Mechanobiology of Diseases: Cancer II
- Lecture 29: Mechanobiology of Diseases: Cancer III
- Lecture 30: Mechanobiology of Diseases: Atherosclerosis & Hypertension

### Week 7

Lecture 31: Mechanobiology of Diseases: Muscular Dystrophy

Lecture 32: Nuclear Mechanotransduction: LINC complex

- Lecture 33: Nuclear Mechanotransduction: LINC complex in cell migration
- Lecture 34: Nuclear Mechanotransduction: Gene regulation
- Lecture 35: Mechanical Forces & DNA damage

#### Week 8

- Lecture 36: Techniques in Mechanobiology: Hydrogels
- Lecture 37: Techniques in Mechanobiology: AFM
- Lecture 38: Techniques in Mechanobiology: Traction Force Microscopy, Trypson Deadhesion & Laser Ablation
- Lecture 39: Techniques in Mechanobiology: Microfabrication
- Lecture 40: Techniques in Mechanobiology: FRE

## **Books and references**

Introduction to Cell mechanics and Mechanobiology, Christopher. R. Jacobs (Garland Science).
 Cellular and biomolecular mechanics and mechanobiology, Editors: Gefen, Amit (Springer).

Duration: 4 weeks Credits: 1

# **Course layout**

**Week 1:** Introduction to nano, Nano-biomimicry, Synthesis of nanomaterials by physical and chemical methods, Synthesis of nanomaterials by biological methods, Characterisation of nanomaterials.

Week 2: DNA nanotechnology, Protein & glyco nanotechnology, Lipid nanotechnology, Bio-nanomachines, Carbon nanotube and its bio-applications.

**Week 3:** Nanomaterials for cancer diagnosis, Nanomaterials for cancer therapy, Nanotechnology in tissue engineering, Nano artificial cells, Nanotechnology in organ printing.

**Week 4:** Nanotechnology in point-of-care diagnostics, Nanopharmacology & drug targeting, Cellular uptake mechanisms of nanomaterials, In vitro methods to study antibacterial and anticancer properties of nanomaterials, Nanotoxicology.

## **Books and references:**

1. Malsch, N.H., "Biomedical Nanotechnology", CRC Press. (2005).

2. Mirkin, C.A. and Niemeyer, C.M., "Nanobiotechnology II: More Concepts and Applications", Wiley-VCH. (2007).

3. Kumar, C. S. S. R., Hormes, J. and Leuschner C., "Nanofabrication Towards Biomedical Applications: Techniques, Tools, Applications, and Impact", WILEY -VCH Verlag GmbH & Co. (2005).

4. Lamprecht, A., "Nanotherapeutics: Drug Delivery Concepts in Nanoscience", Pan Stanford Publishing Pte. Ltd. (2009).

5. Jain, K.K., "The Handbook of Nanomedicine", Humana press. (2008).

# **Biomicrofluidics**

### Duration: 4 weeks Credits: 1

## **Course layout**

- Week 1: Introduction to Biomicrofluidics Engineers' guide to the cell Fluidics in living systems and mechanobiology Pressure driven flows
- Week 2 : Surface tension driven flows Modulating surface tension Lab on a CD Introduction to Electrokinetics
- Week 3 : Microfluidic cell culture On-chip cellular assay techniques Microfluidics for understanding biology
- Week 4 : Organ-on-a-chip Lab-on-a-chip for genetic analysis Microfluidic technology for monoclonal antibody production

## **Books and references**

Nil

## **Fundamentals of Micro and Nanofabrication**

### Duration: 12 weeks Credits: 3

## **Course layout**

Week 1: Introduction to micro-fabrication Week 2: Substrate Week 3: Cleaning Week 4: Additive processing: Doping Week 5: Additive processing: Native Films Week 6: Additive processing: CVD Week 7: Additive processing: PVD Week 8: Lithography 1 Week 9: Lithography 2 Week 10: Subtractive Process: Wet Etching Week 11: Subtractive Process: Dry Etching Week 12: CMP and Packaging

#### **Books and references:**

- 1. The Science and Engineering of Microelectronic Fabrication by Stephen A. Campbell
- 2. VLSI Fabrication Principles: Silicon and Gallium Arsenide by Sorab K. Gandhi
- 3. Introduction to Microelectronic Fabrication by Richard C. Jaeger
- 4. Materials Science of Thin Films by Milton Ohring
- 5. Fundamentals of Microfabrication and Nanotechnology, by Marc J. Madou
- 6. Fundamental Principles of Optical Lithography by Chris Mack
- 7. Electronic Materials by Prof. Dr. Helmut Föll

Duration: 12 weeks Credits: 3

### **Course layout**

#### WEEK: 12 MODULES 40

#### Week 1 Preformulation I

Preformulation I: Physical form (crystal & amorphous), Preformulation I: polymorphism, particle size, shape Preformulation I:, solubility profile (pKa, pH) Preformulation I: Partition coefficient, flow properties

#### Week 2 Preformulation II

Preformulation II: Hydrolysis, oxidation, reduction Preformulation II: Racemisation, Preformulation II: Dissolution, permeability & BCS classification Preformulation II: Polymerization

#### Week 3 Tablets

Tablets: Introduction, classification of tablets. Formulation/preparing of tablets, granulation methods. Tablets: Compression and processing problems. Equipment and tablet tooling, Tablet defects. Tablets: Tablet coating: Types, coating materials, formulation of coating composition, Methods of coating, equipment's employed and defects in coating Tablets: QC of tablets: apparatus, methods, graphical presentations, official and unofficial tests.

#### Week 4 Liquid Orals

Liquid orals: Formulation and manufacturing of solution, syrups and elixirs, Filling and packaging Liquid orals: Emulsions: I Liquid orals: Emulsion II Liquid orals: Suspensions: Formulation, evaluation

#### Week 5 Parenterals I

Parenterals I: Introduction, Preformulation of parenterals Parenterals I: Formulation of Parenterals

#### Week 6 Parenterals I

Parenterals I: Types of Parenteral preparation Parenterals I: Lay out and Production facilities

#### Week 7 Parenterals II

Parenterals II: Pyrogens and pyrogenisity Parenterals II: Quality control of parenterals

### Week 8 Capsules

Capsules: Introduction, Advantages, Disadvantages, Capsule Production, hard Gelatin capsules Capsules: Methods of filling capsules, Soft Gelatin Capsules Capsules: Formulation Consideration, Production of soft gelatin capsules, Non-gelatin soft capsules Capsules: Evaluation of commercial capsules

#### Week 9 Pellets and Ophthalmic Preparations

Pellets: Introduction, formulation, pelletization process, equipment needed Ophthalmic Preparations: Introduction, absorption through eye, formulation considerations Ophthalmic Preparations: formulation of eye drops, lotion, ointments Ophthalmic Preparations: Methods of preparation; labeling, containers Evaluation

| Week 10 | Pharmaceutical Aerosols  |  |  |  |  |
|---------|--|--|--|--|--|
|         | Pharmaceutical Aerosols: Definitions, Advantages, Limitation, Uses                                     |  |  |  |  |
|         | Pharmaceutical Aerosols: Components of aerosols  |  |  |  |  |
|         | Pharmaceutical Aerosols: Components and systems of aerosols  |  |  |  |  |
|         | Pharmaceutical Aerosols: inhalers and Evaluation of Aerosols   |  |  |  |  |
| Week 11 | Cosmetics  |  |  |  |  |
|         | Cosmetics: Introduction, absorption through skin, types of cosmetic preparations, Formulation of tooth |  |  |  |  |
|         | pastes   |  |  |  |  |
|         | Cosmetics: Formulation of lipsticks, shampoos, hair dyes   |  |  |  |  |
|         | Cosmetics: Formulation of cold cream and vanishing cream, sunscreens                                   |  |  |  |  |
| Week 12 | Packaging Materials Science  |  |  |  |  |
|         | Packaging Materials Science: Materials used for packaging of pharmaceutical products,                  |  |  |  |  |
|         | Packaging Materials Science: Factors influencing choice of containers Legal and official requirements, |  |  |  |  |
|         | Stability aspects,   |  |  |  |  |
|         | Packaging Materials Science: Quality control tests of packaging materials                              |  |  |  |  |

## **Books and references:**

- IP/USP/BP
- Semalty et al. Essentials of Pharmaceutical Technology, II edn 2018, reprint 2019, Pharma med Press, Hyderabad
- Niazi SK, Handbook of Preformulation, Informa Health Care, 2007
- Aulton ME (ED), Pharmaceutics: The science of Dosage form design, II edn, Churchill Livingstone, London, 2002.
- Banker GS & Rhodes T, Modern Pharmaceutics, CRC Press.
- Lachman L/Lieberman HA, Kanig JL, The Theory And Practice Of Industrial Pharmacy,4E (2013).
- Qiu Y, Chang Y and Zhang GZ (Exe. Eds), Developing solid oral dosage forms: Pharmaceutical theory and practice, Elsevier, 2009.
- Gibson M. (Ed), Pharmaceutical preformulation and formulation: a practical guide from candidate drug selection to commercial dosage form, II edn, Informa Healthcare.
- Qiu Y, Chang Y and Zhang GZ (Exe. Eds), Developing solid oral dosage forms: Pharmaceutical theory and practice, Elsevier, 2009
- Dressman JB, Karmer J, Pharmaceutical Dissolution Testing, 2005, Taylor & Francis, NY.
- Ansel's Pharmaceutical Dosage Forms and Drug Delivery Systems. Philadelphia: Lipincott Williams and Wilkins.
- Augsburger LL, Hoag SW (Eds), Pharmaceutical Dosage Forms: Tablets, Volume 1, III Edn, Informa healthcare, 2008

# **Mechanical Unit Operations**

### Duration: 12 weeks Credits: 3

## **Course layout**

Week 1: Introduction of Particulate Sizes and Shapes
Week 2: Screening
Week 3: Size Reduction
Week 4: Storage and Conveying of Bulk Solids
Week 5: Size Enlargement
Week 6: Flow past Bluff Bodies
Week 7: Flow Through Packed and Fluidized Beds
Week 8: Filtration
Week 9: Cross Flow Filtration and Membrane Separations
Week 10: Gravity Sedimentation Processes
Week 11: Centrifugal Separations
Week 12: Floatation

### **Books and references:**

1. E. Ortega-Rivas, Unit Operations of Particulate Solids: Theory and Practice, CRC Press, FL, 2012.

2. W.L. McCabe, J. Smith, P. Harriot, Unit Operations of Chemical Engineering, 7th Edition, McGraw Hill, 2005.

3. J.F. Richardson, J.J. Harker, Coulson and Richardson's Chemical Engineering, 2nd Volume, 5th Edition, Butterworth-Heinemann, 2003.

## Nanotechnology in Agriculture

### **Course Duration: 8 weeks**

Credits: 2

## **Course layout**

Week 1: History of agriculture and the role of chemicals in modern agriculture

Week 2: Overview of nanotechnology

Week 3: Application of nanotechnology in modern day agriculture practices I

Week 4: Application of nanotechnology in modern day agriculture practices II

Week 5: Application of nanotechnologies in animal production

Week 6: Nanotechnology and shelf life of agricultural and food products

Week 7: Nanotechnologies for water quality and availability

Week 8: Green nanotechnology and the role of good governance and policies for effective nanotechnology development.

#### **Books and references**

• E-Reference materials will be provided during the course

## Introduction to Biomaterials (being offered as professional elective in regular curriculum)

### **Course Duration: 8 weeks**

## Credits: 2

## **Course layout**

### Week 1:

Introduction to Biomaterials Background history, Properties (Mechanical and Physico-Chemical )

## Week 2:

Mechanical properties resorb ability, biodegradation Resorbability, biodegradation Biofilm

## Week 3:

Biofilm (Material characterization - Analytical instruments Analytical instruments)

## Week 4:

Analytical instruments Analytical instruments Biological responses, compatibility, cytotoxicity; Proteins, Tissue and blood Response Cell-biomaterial interaction

## Week 5:

Animal trials in vivo) Animal trials (metals-types, classifications, applications (metals - properties (metals - properties

## Week 6:

Metals - properties (metals Polymers-types, classifications, applications Polymers Polymers

### Week 7:

Blends/composites Biopolymers Hydrogels Preparation of different morphologies with experiments) Surface modifications with experiments)

### Week8:

Ceramics Drug delivery systems/encapsulation Biomaterials for cardiovascular/pulmonary/ophthalmological applications Biomaterials for urinary/dental/skin applications Sterilization of implants, device failures, unique issues, conclusion.

### **References**:

- 1. Biomaterials Science: An introduction to Materials in Medicine, Edited by Ratner, Hoffman, Schoet and Lemons, Second Edition: Elsevier Academic Press, 2004.
- 2. Comprehensive structural integrity, Vol.9: Bioengineering Editors: Mithe, Ritchie and Karihalo, Elsevier Academic Press, 2003.

## **Additional Reading:**

- 1. Biomaterials Science and Biocompatability, Fredrick H. Silver and David L. Christiansen, Piscataway, Springer, New Jersey.
- 2. Biological Performance of Materials: Fundamentals of Biocompatibility, Janathan Black, Marcel Dekker, Inc., New York and Basel, 1981.
- 3. Basic Cell Culture: A Practical Approach, Edited by J.M. Davis, IRL Press, Oxford University Pres, New York, 1994.

### **Course Duration: 8 weeks**

Credits: 2

# **Course layout**

Week 2: Monitoring wild animals
Week 3: Monitoring & managing habitats
Week 4: Management of wildlife diseases
Week 5: Capturing and restraining wild animals
Week 6: Conservation genetics
Week 7: Ex-situ conservation

Week 1: Introduction, Importance, Threats

Week 8: Management of changes

## **Books and references**

- 1. Pullin, A.S., Conservation Biology. 2002: Cambridge University Press.
- 2. Van Dyke, F., Conservation Biology: Foundations, Concepts, Applications. 2008: Springer Netherlands.
- 3. Selected articles / papers as referred to in the lectures

# **Course Duration: 12 weeks**

Credits: 3

# **Course layout**

Week 1: Introduction to Mathematical Modelling

Week 2: Introduction to Static Networks

Week 3: Network Biology and Applications

Week 4: Reconstruction of Biological Networks

Week 5: Dynamic Modelling of Biological Systems: Introduction, Solving ODEs & Parameter Estimation

Week 6: Evolutionary Algorithms, Guest Lectures on Modelling in Drug Development

Week 7: Constraint-based approaches to Modelling Metabolic Networks

Week 8: Perturbations to Metabolic Networks

Week 9: Elementary Modes, Applications of Constraint-based Modelling

Week 10: Constraint-based Modelling Recap, 13C Metabolic Flux Analysis

Week 11: Modelling Regulation, Host-pathogen interactions, Robustness of Biological Systems

Week 12: Advanced topics: Robustness and Evolvability, Introduction to Synthetic Biology, Perspectives & Challenges

# **Books and references:**

- 1. Raman K (2021) An Introduction to Computational Systems Biology: Systems-Level Modelling of Cellular Networks. 1/e ISBN 9781138597327 (Chapman and Hall/CRC)
- 2. Voit E (2012) A First Course in Systems Biology. Garland Science, 1/e. ISBN 081534467
- 3. Klipp E (2009) Systems biology: a textbook. Wiley-VCH, 1/e. ISBN 9783527318742
- 4. Newman MEJ (2011) Networks: an introduction. Oxford Univ. Press. ISBN 9780199206650

## **Drug Delivery: Principles and Engineering**

Duration: 12 weeks Credits: 3

Prerequisites: A course in biochemistry, molecular biology, anatomy is recommended

### **Course layout**

Week 1: Pharmacokinetics: Bioavailability, Elimination, Therapeutic index

Week 2: Prodrugs, Controlled release

Week 3: Polymers: Synthesis, properties, characterization, crystallinity and amorphousness

- Week 4: Biopolymers: Natural and Synthetic, biocompatibility, Biodegradation, commonly used biopolymers
- Week 5: Polymer-Drug conjugates, PEGylation
- Week 6: Diffusion controlled systems, Ficks laws, Reservoir systems, non-erodible matrix systems, Bio-erodible systems
- Week 7: Hydrogels: Physical or chemical, pore-size calculation, in-situ crosslinking

Week 8: Nano and Micro-particles: Dendrimers, Liposomes, Micelles

Week 9: Metal and polymeric particles, effect of particle shape, charge and elasticity

Week 10: Protein Adsorption and tissue engineering, Drug delivery in tissue engineering

- Week 11: Implant associated infections, Route specific delivery: Oral, Subcutaneous, Intramuscular, transdermal, inhalation, intravenous
- Week 12: Vaccines, Cancer vaccines, Cell and gene delivery, Smart responsive drug delivery, Targeted drug delivery, Nanotoxicology and market translation

## **Books and references:**

- 1. Drug Delivery: Engineering Principles for Drug Therapy, W. Mark Saltzman, Oxford University Press, 2001.
- 2. Drug Delivery: Fundamentals and Applications, Anya M. Hillery and Kinam Park, 2nd Edition, CRC

Press, 2016.

Duration: 12 weeks Credits: 3

Prerequisites: Nil

## **Course layout**

**Week 1**: Proteogenomics overview- Part I, Proteogenomics overview- Part II, Introduction to Genomics- Part I: Gene sequencing and mutations Introduction to Genomics-Part II: Sequence alignment, Introduction to Genomics-Part III: Transcriptome, SL1: Advancement in Cancer Genomics, SL2: Advancement in Cancer Genomics

Week 2: Introduction to Genomics IV: Epigenome, Introduction to Genomics: cBioPortal, Genotype, Gene expression & Phenotype - Part I, Genotype, Gene expression & Phenotype Part II, An overview of NGS technology, SH1: NGS-Sequencing by synthesis, SH2: NGS Sequencing by synthesis

Week 3: Introduction to Proteomics, Proteomics: Sample Prep & Protein Quantification, Proteomics: Sample Prep & Protein Quantification (Hands-on), Introduction to MS-based Proteomics- Part I, Introduction to MS-based Proteomics- Part II, SL 3: Applications of NGS – Ion Torrent, SL4: Applications of NGS – Ion Torrent

**Week 4**: Introduction to MS-based Proteomics- Part I (Hands-on), Introduction to MS-based Proteomics- Part II (Hands-on), Data analysis: Normalization, Data analysis: Batch Correction and Missing values, Data analysis: Statistical Tests, SH3: NGS- Ion Torrent, SH4: NGS- Ion Torrent

**Week 5**: Machine learning and Clustering, Hypothesis testing, ProTIGY- Part I, ProTIGY- Part II, Proteogenomics approach to unravel proteoforms, SL5: Genomic Analysis using Droplet PCR, SL6: Genomic Analysis using Droplet PCR

Week 6: Workflow to Automated Data Processing, Introduction to Fire Cloud, Fire Cloud and Data Model, Bioinformatics solutions for 'Big Data' Analysis-Part I, Bioinformatics solutions for 'Big Data' Analysis-Part II, SH5: Genomic Analysis using Droplet PCR, SH6: Genomic Analysis using Droplet PCR

Week 7: Data Science infrastructure management- Part I, Data Science infrastructure management-Part II, Data Science infrastructure management- Part III, DIA-SWATH Atlas Part I, DIA-SWATH Atlas-Part II, SL7: Introduction to Targeted Proteomics, SH7: Data Analysis using Skyline

**Week 8**: Human Protein Atlas-Part I Clinical, Human Protein Atlas-Part II, Affinity based proteomics & HPA, Clinical Considerations for OMICS-Part I, Considerations for OMICS- Part II, SL8: Proteomics: PTMs, SL9: Clinical Proteomics

Week 9: Introduction to Proteogenomics-Part I, Introduction to Proteogenomics-Part II, Sequence centric proteogenomics, Gene Variant Analysis, Proteomics in Clinical studies, SH8: ProTIGY

Week 10: Supervised Machine learning- Predictive Analysis Part I, Supervised Machine learning-Predictive Analysis Part II, Supervised Machine learning- Marker Selection, Gene Set Analysis using WebGestalt- Part I, Gene Set Analysis using WebGestalt- Part II, SH9: Supervised Machine Learning

Week 11: Biological Network Analysis- Part I, Biological Network Analysis- Part II, Mutation and Signaling - Part I, Mutation and Signaling- Part II, Pathway Enrichment, SH10: Pathway Enrichment and Network Analysis

Week 12: Gene Set Enrichment Analysis (GSEA), Pathway enrichment: GSEA, Linked Omics, Linked Omics (Hands-on), Proteogenomics Conclusions, SL10: Topics in Proteogenomics Malaria and Cancer case study

## **Books and references:**

1. Proteomics: A Cold Spring Harbor Laboratory Course Manual, A.J. Link and J. LaBaer, Cold Spring Harbor Laboratory Press, 2009. Selected papers from scientific journals

Duration: 12 weeks Credits: 3

Prerequisites: A background degree in Science or Technology is preferable. Students who enroll for this course may also benefit from the course "Patent Drafting for Beginners"

## **Course layout**

**Week 1**: Introduction to the Indian Patent System Patent Laws as Concepts; Understanding the Patents Act, 1970; Understanding the Patents Rules, 2003; Preliminary Sections; Preliminary Rules; What's New in the Patents (Amendment) Rules, 2016; Easy way to read the Patents Act and Rules.

**Week 2**: Patentability of Inventions Statutory Exceptions to Patentability; Novelty and Anticipation; Inventive Step; Capable of Industrial Application; Person Skilled in the Art.

**Week 3**: Patent Specification Provisional and Complete Specifications; Structure of a Patent Specification—Title, Abstract, Description, Claims, etc.; Reading a Patent Specification—Fair basis, Enabling Disclosure, Definiteness, Priority; Introduction to Patent Drafting.

**Week 4**: Patent Prosecution: Patent Applications Patent Application—Who Can Apply, True and First Inventor, How to Make a Patent Application, What to include in a Patent Application, Types of Patent Applications, Patents of Addition, Dating of Application.

**Week 5**: Patent Prosecution: Publication and Examination - I Publication of Application; Request for Examination; Examination of Application—First Examination Report.

**Week 6**: Patent Prosecution: Publication and Examination – II Expedited Examination of Application; Search for Anticipation—Procedure, withdrawal of Application; Consideration of Report of Examiner

Week 7: Patent Prosecution: Powers of Controller Powers of Controller—Examination Stage, Consideration of report by examiner, Refuse or Amend Applications, Division of Applications, Dating of Application, Anticipation, Potential Infringement; Putting Applications in Order; Amendments during Prosecution

**Week 8**: Patent Prosecution: Opposition Pre-grant opposition; Post-grant opposition; Wrongful obtaining of invention; Mention of Inventor; Opposition in General.

Week 9: Patent Prosecution: Practice at the Patent Office- I Secrecy Provisions; Grant of Patents; Rights Conferred by Grant; Rights of Co-Owners; Term of Patent; Restoration of Lapsed Patents;

Week 10: Patent Office and Patent Prosecution, Surrender; Revocation—Grounds for Revocation; Register of Patents, Patent Office and its Establishment; Patent Agents; Use and Acquisition by Government; Penalties.

Week 11: Compulsory Licensing Compulsory Licensing—Working of Patents, Grounds for Grant of Compulsory License, Revocation; Patent Licensing;

Week 12: Patent Enforcement, International Arrangements and Other Miscellaneous Provisions Intellectual Property Appellate Board; Declaratory Suits, Infringement Suits; International ApplicationConvention Application, PCT Application, Application Designating India, Multiple Priorities; PCT Timeline; Fees—Application, In Relation to Grant of Patents; Timelines, Application, Examination, Publication etc.

## **Books and references**

- 1. Feroz Ali, The Law of Patents, LexisNexis Ronald D. Slusky, Invention Analysis and Claiming A Patent Lawyer's Guide, Second Edition, American Bar Association, 2012.
- 2. Feroz Ali, the Touchstone Effect The Impact of Pre-grant Opposition on Patents, LexisNexis, 2009.

Duration: 8 weeks Credits: 2

## Prerequisites: Prior knowledge of biochemistry, bioinformatics

## **Course layout**

Week 1: Introduction to drug discovery

Week 2: Structure and property

Week 3: ADME-rules

Week 4: Force field/MM/QM

Week 5: Boundary conditions/Conformation

Week 6: QSAR/ Pharmacophore

Week 7: Enzymes/proteins structures/docking

Week 8: PK/PD

## **Books and references:**

- 1. Voit E (2012) A First Course in Systems Biology. Garland Science, 1/e. ISBN 0815344678.
- 2. Klipp E (2009) Systems biology: a textbook. Wiley-VCH, 1/e. ISBN 9783527318742.
- 3. Newman MEJ (2011) Networks: an introduction. Oxford Univ. Press. ISBN 9780199206650.

Duration: 8 weeks Credits: 2

Prerequisites: The target audiences of this course are required to have a basic introduction to biology.

**Course layout** 

Week 1: Basics of Proteins and Proteomics Lecture 1: Introduction to amino acids Lecture 2: Introduction to Proteins Lecture 3: Protein folding & misfolding Lecture 4: Introduction to Proteomics Lecture 5: Lab session – Protein-protein interaction using label-free biosensors

Week 2: Gel-based proteomics

Lecture 6: Sample preparation and pre-analytical factors

Lecture 7: Sample preparation: pre-analytical factors (contd.)

Lecture 8: Sample preparation: Protein extraction and quantification

Lecture 9: One-dimensional electrophoresis

Lecture 10: Introduction to 2-DE

Week 3: Two-dimensional gel electrophoresis (2-DE)

Lecture 11: 2-DE: Second dimension, staining & destaining

Lecture 12: 2-DE: Gel analysis

Lecture 13: 2-DE Applications

Lecture 14: 2-DE Applications (contd.) & Challenges

Lecture 15: Lab session - Protein/peptide pre-fractionation using OFFGEL FRACTIONATOR & data analysis

Week 4: Difference in gel electrophoresis (DIGE) & Systems Biology Lecture 16: 2D-DIGE: Basics Lecture 17: 2D-DIGE: Data analysis Lecture 18: 2D-DIGE: Applications Lecture 19: Systems biology and proteomics – I Lecture 20: Systems biology and proteomics – II

Week 5: Basics of mass spectrometry

Lecture 21: Fundamentals of mass spectrometry

Lecture 22: Chromatography technologies

Lecture 23: Liquid chromatography

Lecture 24: Mass spectrometry: Ionization sources

Lecture 25: Mass spectrometry: Mass analyzers

Week 6: Basics of mass spectrometry and sample preparation

Lecture 26: MALDI sample preparation and analysis

Lecture 27: Hybrid mass spectrometry configurations

Lecture 28: Lab session - Demonstration of Q-TOF MS technology

Lecture 29: In-gel & in-solution digestion

Lecture 30: Lab session - Sample preparation: tissue sample preservation technology

Week 7: Quantitative proteomics Lecture 31: Introduction to quantitative proteomics Lecture 32: SILAC: In vivo labelling Lecture 33: iTRAQ: In vitro labelling Lecture 34: TMT: In vitro labelling Lecture 35: Quantitative proteomics data analysis

Week 8: Advancement in Proteomics Lecture 36: Proteomics applications Lecture 37: Challenges in proteomics Lecture 38: OMICS and translational research Lecture 39: Lab session – Targeted proteomics using triple quadrupole mass spectrometry Lecture 40: Lab session – Targeted proteomics: multiple reaction monitoring

### **Books and references:**

Nil

# Introduction to cell culture Technology

Duration: 8 weeks Credits: 2

## **Course layout**

Week 1: Introduction & biology of cultured cells

Week 2: Equipments, aseptic techniques, safety protocols

Week 3: Culture vessels & media development

Week 4: Serum-free medium development & sterilization

Week 5: Primary culture, secondary culture, cloning & selection

**Week 6:** Cell separation, characterization, differentiation & transformation

Week 7: Contamination, cryo-preservation & cyto-toxicity

Week 8: Organo-typic culture & specialized cell culture techniques

# **BOOKS AND REFERENCES: NIL**

## Maternal Infant Young Child Nutrition

**Course Duration: 12 Weeks Credits: 3** 

**Course layout** 

- Week 1: Maternal Infant Young Child Nutrition Introduction
- Week 2: Science of nutrition
- Week 3: Types of malnutrition and hidden hunger
- Week 4: Importance of first 1,000 days
- Week 5: Science of Breastfeeding

Week 6: Cross cradle hold and 45 points of breastfeeding counselling.

- Week 7: Other breastfeeding holds
- Week 8: Manual expression, storage and feeding of the expressed breastmilk. Nipple/Breast Conditions.
- Week 9: New born care and Kangaroo mother care
- Week 10: Complementary feeding
- Week 11: Maternal Nutrition-pre-pregnancy pregnancy-lactating mothers Adolescent Nutrition
- Week 12: Assessment of anthropometric measurement and growth charts Percentile & Z Score

#### **Books and references:**

WHO Growth Charts

# Optical Spectroscopy and Microscopy: Fundamentals of Optical Measurements and Instrumentation

**Course Duration: 12 weeks** 

Credits: 3

# **Course layout**

**Week 1**: Essential Quantum Mechanics: Uncertainity Principle, Probabilistic nature of measurement, postulates of qmech, Stern Gerlach equivalent in light, Photon picture (PMT response), Linear Vector Space.

**Week 2:** Time dependent perturbation theory, Fermi Golden Rule, Transition probability in light matter interaction, Beer Lambert relation, Einestin's phenomenolgical treatment, A and B coefficients, Spontaneous emission, Origins of fluorescence

Week 3: Nature of Fluorescence, Emisson spectrum, Absorption spectrum, Anisotropy, Life time, FRET

Week 4: Second quantaisation, creation and anhilation operators, Fock states, light matter interaction in Feynman digrams

Week 5: Spontaneous emission orgin, Stimulated Emission origin dependence through Fock states

Week 6: Laser emission, two state, three state and four state laser systems

Week 7: Real world lasers, Charecteristics of laser emission, thershold behavior, Laser gain equation, CW operation, Pulsed lasers, Qswitching, mode locking, Saturable absorber

**Week 8:** Laser induced fluorescence, optical components (lenses, mirrors, gratings, prisms) and their working principles, Interference filters, dichroic filters, efficiency calculations for SNR improvement, aligning an optical equipment.

Week 9: Intro to optical hardware, common opto-mechanical assemblies, setting up a simple laser based spectrometer using gratings in lab, calibration and acquisition of fluorescein spectra.

**Week 10:** Principles of photo detection, QE, Dynamic range shot noise, photodetectors – PMTs, photodiodes, photo resistors, understanding common metrics and specs. Detection electronics – preamps, A2Ds

Week 11: Area detectors, CCDs, emCCDs, sCMOS, comparison, read noise, speed and other sensor charecteristics. Theory of Image formations – widefiled microscopy, bright field, phase contrast, DIC and fluorescence microscopy

Week 12: Scanning system: Principles of scanning system, Gaussian light progation and focussing, optical resolution, deinition in xy and z. Measurement and charecetrisation in lab. Scanning as time averaged focus, optical hinges, imaging of hinges, Confocal microscope

## **Books and references:**

1. Optical Spectroscopy (Demtroder), Quantum Electronics - Yariv, Building Scientific Apparatus - WJ Moore

## **Course Duration: 4 Weeks**

## Credits: 1

## **Course layout**

Week 1: Fundamentals of central dogma (DNA, RNA and proteins; mutations), Chromosome structure and function (organization; structure- function relationship; chromosome abnormalities).

**Week 2**: Genes in pedigree (Mendelian pedigree patterns, complications to pedigree patterns), DNA cloning and hybridization techniques (vector-based cloning; nuclei acid hybridizations; PCR-based DNA analyses)

Week 3: Mutation and instability of human DNA (mutation and polymorphism; pathogenic mutations, repeat expansions), Molecular pathology (types of mutations; animal models for human disease)

Week 4: Identifying human disease genes (functional cloning versus positional cloning; mutation screening), Complex diseases; The Human Genome and HapMap projects

#### **Books and References:**

Human Molecular Genetics 4 Tom Strachan, Andrew P. Read Garland Science/Taylor & Francis Group, 2011

# Demystifying the Brain

## **Course Duration: 4 weeks**

Credits: 1

## **Course layout**

Week 1: History of neuroscience, Brain through evolution

Week 2: Neurons and neural signaling Networks that learn

Week 3: Organization of the nervous system Maps in the brain

Week 4: Memories and holograms, Emotions in the brain, Theories of Consciousness

- 1. Demystifying the brain, ebook on NPTEL.
- 2. Valentino Braitenberg, Vehicles.
- 3. VS Ramachandran, Phantoms in the Brain. Joseph LeDoux, Emotional Brain

Course Duration: 8 weeks Credits: 2

## **Course layout**

Week 1: Introduction to Biomaterials Background history History Properties (Mechanical and Physico chemical) Properties (Mechanical and Physico-chemical)

Week 2: Mechanical properties Mechanical properties Resorbability, biodegradation Resorbability, biodegradation Biofilm

Week 3: Biofilm Biofil m Material characterization - Analytical instruments Analytical instruments

Week 4: Analytical instruments Analytical instruments Biological responses, compatibility, cytotoxicity Proteins, Tissue and blood Response Cell-biomaterial interaction

Week 5: Animal trials (in vivo) Animal trials Metals-types, classifications, applications Metals - properties

Week 6: Metals - properties Metals Polymers-types, classifications, applications Polymers

**Week 7:** Blends/composites Biopolymer s Hydrogels Preparation of different morphologies (with experiments) Surface modifications (with experiments)

Week8:CeramicsDrugdeliverysystems/encapsulationBiomaterialsforcardiovascular/pulmonary/ophthalmologicalapplicationsBiomaterialsforurinary/dental/skinapplicationsSterilization of implants, device failures, unique issues, conclusion

#### **Books and references:**

NIL

# Forests and Their Management

## Course Duration: 12 weeks Credits: 3

## **Course layout**

Week 1: Introduction

- Week 2: Basics of silviculture
- Week 3: Forest soils
- Week 4: Forest mensuration
- Week 5: Forest surveying
- Week 6: Forest protection
- Week 7: Silvicultural management I
- Week 8: Silvicultural management II
- Week 9: Logging and yield
- Week 10: Silvicultural practices
- Week 11: Newer trends in forestry
- Week 12: Revision

- 1. Principles and practices of Silviculture by S. S. Bist
- 2. Forest soils by Wilde

## **Course Duration: 4 weeks**

Credits: 1

# **Course layout**

Week 1: Fundamentals of electrochemistry with special references to bioelectrochemistry

Week 2: Electrodes & potentiometry

Week 3: Redox titrations

Week 4: Electro-analytical techniques

## **Books and references:**

1. Quantitative chemical analysis by Daniel C Harris

2. D. Bioelectrochemistry: Fundamentals, Applications and Recent Developments Richard C. Alkire (Editor), Dieter M. Kolb (Editor), Jacek Laskowski (Editor), Phil N. Ross (Series Editor).

#### **Course Duration: 4 weeks**

## Credits: 1

## **Course layout**

Week 1: Introduction: Life cycle of an angiosperm plant, Plant growth and development, Embryonic and postembryonic development, Characteristics of plant development

**Week 2**: Molecular Genetics of Plant Development: Generation and characterization of developmental mutants, studying temporal and spatial expression pattern of developmental regulators, Functional genomics, Genetic manipulation of plant for studying development

Week 3: Root development: Organization and maintenance of root apical meristem, radial patterning during vascular development, Root branching; lateral root development

Week 4: Shoot development: Organization and maintenance of shoot apical meristem, Organogenesis and organ polarity, Floral transition, Floral organ patterning and determinacy, Cell-to-cell communication during development.

- 1. Leyser, O. and Day, S. Mechanisms in plant development. John Wiley & Sons. 2009.
- 2. Howell, S.H. Molecular genetics of plant development. Cambridge University Press. 1998.
- 3. Taiz, L. and Zeiger, E. Plant Physiology. Sinauer Associates. 2010 5th Eds.
- 1. Raven, P.H. Evert, R.F. and Eichhorn, S.E. Biology of plants. Macmillan. 2005 8th Eds.

## **Conservation Economics**

## **Course Duration: 12 weeks**

Credits: 3

## **Course layout**

Week 1 What is Economics?

Week 2 What is Conservation?

Week 3 Modern impacts necessitating conservation

Week 4 Threats to wildlife

Week 5 How can Economics help?

Week 6 Markets: Places where Economics works

Week 7 Markets, welfare and conservation

Week 8 Public sector and conservation

Week 9 Industrial organization and conservation

Week 10 Labour market economics and conservation

Week 11 Practical issues in Economics and Conservation

Week 12 Case Studies

- 1. Economics, Krugman and Wells
- 2. Economics, Hubbard & O'Brien
- 3. Principles of Economics, N. Gregory Mankiw
- 4. Basic Economics, Thomas Sowell

## Soft Nano Technology

#### **Course Duration: 8 weeks**

Credits: 2

#### **Course layout**

Week 1: Introduction to Patterning of Thin Films Application of Nano Patterned Films and Surfaces Basic Concepts of Wetting: Cassie and Wenzel Regimes Basic Concepts of Surface Tension

Week 2: Different Nano Fabrication Regimes including self-assembly Micelle formation Introduction to Photo Lithography

Week 3: Discussion on Photo Lithography: Photo Resists Spin Coating Exposure Development

Week 4: Nano Imprint Lithography

Week 5: Soft Lithography: Introduction Different Techniques

Week 6: Soft Lithography Techniques

Week 7: Basic Concepts of Atomic Force Microscopy

Week 8: Different Imaging Modes of Atomic Force Microscopy

#### **Books and references:**

1. Alternative Lithography", C. M. Sotomayor Torres (Ed.), Kluwer Academic Press, 2003.

2. Creating Micro and Nano Patterns on Polymeric Materials", A del Campo and E. Arzt (Ed), Wiley, 2011.

3. "Micro Fluidics and Micro Scale Transport Process", Suman Chakraborty (Ed), CRC Press, 2013

## **Understanding Design**

#### **Course Duration: 4 weeks**

#### Credits: 1

#### **Course layout**

Week 1:

Module 1- An Introduction to Design, Module 2- Users and Context

Week 2: Module 3-Design and Society, Module 4 - Design and Sustainability

#### Week 3:

Module 5 - Design and Industry, Module 6 - Design and collaboration

Week 4: Module 7 - Innovation by Design

- 1. Ansell, C & Torfing J (eds) (2014). Public Innovation through Collaboration and Design.London and New York:
- 2. Routledge. Antonelli, Paola (2005). Humble Masterpieces: everyday marvels of Design. Harper Collins Publishers.
- 3. Baxter, Mike (1995). Product Design. London Glasgow New York:
- 4. Chapman & Hall. Brown, Dan M (2013). Designing Together. New Riders.
- 5. Doordan, Dennis (ed) (2000). Design History: An Anthology. Cambridge, London: MIT
- 6. Press. Heskett, John (2002). Design: a very short introduction. Oxford University Press.
- 7. Geist, Valerius (1978). Life Strategies, Human Evolution, and Environmental Design: towards a biological theory of health. New York, Heidelberg, Berlin: Springer-Verag
- 8. Lawson, Brian (2006). How Designer's Think: The design process demystified.
- 9. Routledge. Highmore, Ben (ed) (1975). The Design Culture Reader. London and New York:
- 10. Routledge. Kepes, Gyorgy (ed) (1966). The Man-Made Object. Studio Vista
- 11. London. Norman, Don (2013). The Design of Everyday Things. Hachette UK.
- 12. Papanek, Victor J (1984). Design for the Real World: Human Ecology and Social Change. Academy Chicago. Essi Salonen Designing Collaboration Link
- 13. Gupta, Anil K, Grassroots Innovation: Minds On The Margin Are Not Marginal Minds
- 14. Link Brown Tim, Change by Design: How Design Thinking Transforms Organizations
- 15. and Inspires Innovation Link D'Source, IDC, IITBombay: http://www.dsource.in/

## **Design, Technology and Innovation**

## **Course Duration: 8 weeks**

## Credits: 2

## **Course layout**

Week 1: i. Jaipur Foot - A classic innovation by Prof. B. K. Chakravarthy
ii.User Centered Helmet Design by Prof. B. K. Chakravarthy
Week 2: Challenges of Reaching a Million Users by Prof. Chetan Solanki and Prof
Jayendran V
Week 3: i. Technology to Solution by Prof. Ramesh Singh ii. Collaborative Excellence by Prof. B. Ravi & Prof. B.
K. Chakravarthy
Week 4: Collaborative Innovation Methods by Prof B. K. Chakravarthy
Week 5: Learnings from Grassroot Innovation by Prof. Anil Gupta
Week 6: Systemic Approach to Biomed Innovations by Prof. B. Ravi
Week 7: Research to Innovation by Prof. Amaresh Chakrabarti
Week 8 : Smartcane for the Blind- A Success Story by Prof. P. V. Madhusudhan Rao

## **Books and references:**

Nil

# **Emotional Intelligence**

#### **Course Duration: 8 weeks**

Credits: 2

# **Course layout**

Week 1: Introduction to emotion, intelligence & wisdom

Week 2: Concept, theory, measurement and applications of intelligence

Week 3: Emotional intelligence: concept, theory and measurements

Week 4: Correlates of emotional intelligence

Week 5: Emotional intelligence, culture, schooling and happiness

- Week 6: For enhancing emotional intelligence EQ mapping
- Week 7: Managing stress, suicide prevention, through emotional intelligence, spirituality and meditation

Week 8: Application of emotional intelligence at family, school and workplace

#### **Books and references:**

NIL

# **Exploring Survey Data on Health Care**

**Course Duration: 8 weeks** 

Credits: 2

## **Course layout**

Week 1: Introduction to Health Care Data

Week 2: Preparation for Field Survey on Health Care

Week 3: Testing of Sample Data

Week 4: Handling of Data Software

Week 5: Survey Data using STATA

Week 6: Analysis of Data

Week 7: Panel Survey Data

Week 8: Policy Evaluation of Health Care

- 1. William H Greene: Econometric Analysis, Pearson 8th Edn.
- 2. Freedman & Pisani & Purves: Statistics 4th Edn
- 3. Cameron & Trivedi: Micro econometrics using STATA, Revised Edn.
- 4. Damodar Gujarati, Econometrics: By example, 2nd edition
- 5. Wooldridge: Econometric Analysis of Cross-section and Panel Data, MIT Press
- 6. Consoli, S; Recupero, D; Petkovic, M (2019), Data Science for Healthcare: Methodologies and applications
- 7. Drummond, M; Sculpher, M; Torrance, G (2005), Methods for the Economic
- 8. Evaluation of Health Care Programmes Other Readings during lecture ppts.

#### **Material and Energy Balances**

#### **Course Duration: 12 weeks**

Credits: 3

#### **Course layout**

Week 1: Introduction; Units and dimensions; Basic terminologies

Week 2: Fundamentals of Material Balances; Material Balances for Single Units Without Reactions

Week 3: Material Balances for Multiple Units Without Reactions; Material Balances for Reactive Processes

Week 4: Material Balances for Reactive Processes; Combustion Reactions

Week 5: Material Balances for Systems with Recycle, Bypass, and Purge

Week 6: Energy Balance Terminologies; Introduction to Energy Balances

Week 7: Mechanical Energy Balances; Objectives and Procedures for Energy Balances

Week 8: Energy Balances on Nonreactive Processes without Phase Change

Week 9: Energy Balances on Nonreactive Processes with Phase Change

Week 10: Mixing and Solutions; Fundamentals for Energy Balances on Reactive Processes

Week 11: Energy Balances on Reactive Processes

Week 12: Material and Energy Balances for Unsteady State Processes

- 1. David M. Hummable and James B. Riggs, Basic Principles and Calculations in Chemical Engineering, 7th Edition, Publisher: Prentice Hall India
- 2. Richard M. Felder and Ronald W. Rousseau, Elementary Principles of Chemical Processes, 3rd edition, Publisher: John Wiley & Sons
- 3. Pauline Doran, Bioprocess Engineering Principles, 2nd Edition, Publisher: Academic Press Ann Saterbak, Ka-Yiu San, Larry V. McIntire, Bioengineering Fundamentals, Publisher: Pearson

## **Course Duration: 8 weeks**

Credits: 2

## **Course layout**

Week 1: Introduction to the course

Week 2: Design of batch bioreactors

Week 3: Design of fed-batch bioreactors

Week 4: Design of continuous mode of bioreactors

Week 5: Mass transfer in bioreactors, Rheology of fermentation broths, Heterogeneous reactions in bioprocesses

Week 6: Heterogeneous reactions in bioprocesses (contd.), Heat transfer in bioreactors

Week 7: Heat transfer in bioreactors (contd.) Scale-up of bioreactors: criteria for scale-up, scale-up parameters

Week 8: Scale-up of bioreactors (contd.), non-ideal reactors: design and analysis

- 1. Michael L. Shuler and Fikret Kargi, Bioprocess Engineering: Basic Concepts, Prentice Hall, 1992
- 2. James M. Lee, Biochemical Engineering, Prentice Hall, 1992
- 3. Pauline Doran, Bioprocess Engineering Principles, 2nd Edition, Academic Press 2012
- 4. James E. Bailey and David F. Ollis, Biochemical Engineering Fundamentals, McGraw Hill 5. 1986.
- 6. S. Liu, Bioprocess Engineering: Kinetics, Biosystems, Sustainability, and Reactor Design, Elsevier, 2016
- 7. Octave Levenspiel, Chemical Reaction Engineering, Wiley 2016.

## Waste to Energy Conversion

#### **Course Duration: 8 weeks**

Credits: 2

#### **Course layout**

Week 1 - Introduction, characterization of wastes.

Week 2 - Energy production form wastes through incineration, energy production through gasification of wastes.

Week 3 - Energy production through pyrolysis and gasification of wastes, syngas utilization.

- Week 4 Densification of solids, efficiency improvement of power plant and energy production from waste plastics.
- Week 5 Energy production from waste plastics, gas cleanup.
- Week 6 Energy production from organic wastes through anaerobic digestion and fermentation, introduction to microbial fuel cells.
- Week 7 Energy production from wastes through fermentation and transesterification.

Week 8 - Cultivation of algal biomass from wastewater and energy production from algae.

- 1. Rogoff, M.J. and Screve, F., "Waste-to-Energy: Technologies and Project Implementation", Elsevier Store.
- 2. Young G.C., "Municipal Solid Waste to Energy Conversion processes", John Wiley and Sons.
- 3. Harker, J.H. and Backhusrt, J.R., "Fuel and Energy", Academic Press Inc.
- 4. EL-Halwagi, M.M., "Biogas Technology- Transfer and Diffusion", Elsevier Applied Science.
- Hall, D.O. and Overeed, R.P.," Biomass Renewable Energy", John Willy and Sons. Mondal, P. and Dalai, A.K. eds., 2017. Sustainable Utilization of Natural Resources. CRC Press.

#### **Course Duration: 12 weeks**

Credits: 3

#### **Course layout**

Week 1: Introduction to Water Pollution and Control

Week 2: Pre-treatment & Physical treatment: Flow equalization & Aeration

Week 3: Pre-treatment & Physical treatment: Coagulation and Flocculation

Week 4: Setting and Sedimentation & Settling Chamber Design

Week 5: Filtration & Filtration System Design

Week 6: Wastewater treatment by Adsorption & Ion Exchange

Week 7: Wastewater treatment by Membrane Based Technologies

Week 8: Advanced Oxidation Processes: Introduction

Week 9: Advanced Oxidation Processes: Fenton and catalytic treatment

Week 10: Advanced Oxidation Processes: Photo-induced processes

Week 11: Advanced Oxidation Processes: Sono- and Electro-chemical Treatment

Week 12: Case studies on wastewater treatment in various process, chemical and allied industries

- 1. Weber, W.J., "Physico-chemical Processes", Wiley Interscience, 1983.
- 2. Eckenfelder W.W., "Industrial Water Pollution Control", 2nd Ed., McGraw Hill, 1999.
- 3. Tchobanoglous G., Burton F.L., Stensel H.D., "Metcalf and Eddy Inc.- Waste Water Engineering Treatment and Reuse", Tata McGraw-Hill, 2017
- 4. Arceivala S.J. and Asolekar S.R., "Wastewater Treatment for Pollution Control and Reuse", 3rd Ed., Tata McGraw Hill, 2007.
- 5. Sincero A.P. and Sincero G.A., "Environmental Engineering A Design Approach", Prentice Hall, 1996.
- 6. R.L.Droste, "Theory and Practice of Water and Wastewater Treatment", John Wiley, 1997.
- 7. S. Vigneswaran and C. Visvanathan, "Water Treatment Processes: Simple Options", CRC Press, 1995.

## Renewable Energy Engineering: Solar, Wind and Biomass Energy Systems

**Course Duration: 12 weeks** 

Credits: 3

#### **Course layout**

Week 1: Solar Energy: Basics and Concepts

Week 2: Non-Concentrating Solar Collectors

Week 3: Non-Concentrating Solar Collectors: Practice Problems

Week 4: Concentrating Solar Collectors

Week 5: Storage Systems

Week 6: Biomass types and characterization

Week 7: Biochemical conversion processes

Week 8: Biochemical conversion processes (Contd.)

Week 9: Bioconversion of substrates into alcohol and thermo-chemical conversion of Biomass

Week 10: Bioconversion of substrates into alcohol and thermo-chemical conversion of biomass (Contd.)

Week 11: Wind Energy: Basics: Turbine terms, types and theories

Week 12: Characteristics and Power Generation from Wind Energy

- 1. Sukhatme S. P., Nayak J. K., Solar Energy: Principles of thermal Collection and Storage, 3 rd Ed., Tata McGraw-Hill Education Pvt. Ltd 2008.
- 2. Twidell, J. and Tony W., Renewable Energy Resources, 2 nd Edition, Taylor & Francis 2006.
- 3. Khan B. H., Non-Conventional Energy Resources, 2 nd Edition, Tata McGraw-Hill Education Pvt. Ltd. 2009.
- 4. Prabir Basu, Biomass Gasification, Pyrolysis and Torrefaction, Academic Press, Elsevier, 2013.

Course Duration: 12 weeks Credits: 3

# **Course layout**

Week 1: Introduction: World energy scenario, consumption pattern, fossil fuel depletion and environmental issues

**Week 2**: Biomass: Availability and abundance, photosynthesis, composition and energy potential, virgin biomass production and selection, waste biomass (municipal, industrial, agricultural and forestry) vailability, abundance and potential, biomass as energy resources: dedicated energy crops, annual crops (maize, orghum sugar beet, hemp), perennial herbaceous crops (sugarcane, switchgrass, miscanthus), short rotation woody crops (poplar, willow), oil crops and their biorefinery potential, microalgae as feedstock for biofuels and biochemical, enhancing biomass properties for biofuels, challenges in conversion

Week 3: Biorefinery: Basic concept, types of biorefineries, biorefinery feedstocks and properties, economics

Week 4: Biomass Pretreatment: Barriers in lignocellulosic biomass conversion, pretreatment technologies such as acid, alkali, autohydrolysis, hybrid methods, role of pretreatment in the biorefinery concept

Week 5: Physical and Thermal Conversion Processes: Types, fundamentals, equipment's and applications; thermal conversion products, commercial success stories

Week 6: Microbial Conversion Process: Types, fundamentals, equipment's and applications, products, commercial success stories

Week 7: Biodiesel: Diesel from vegetable oils, microalgae and syngas; transesterification; FT process, catalysts; biodiesel purification, fuel properties

Week 8: Biooil and Biochar: Factors affecting biooil, biochar production, fuel properties, biooil upgradation

**Week 9**: Bioethanol and Biobutanol: Corn ethanol, lignocellulosic ethanol, microorganisms for fermentation, current industrial ethanol production technology, cellulases and their role in hydrolysis, concepts of SSF and CBP, advanced fermentation technologies, ABE fermentation pathway and kinetics, product recovery technologies

Week 10: Hydrogen, Methane and Methanol: Biohydrogen generation, metabolic basics, feedstocks, dark fermentation by strict anaerobes, facultative anaerobes, thermophilic microorganisms, integration of biohydrogen with fuel cell; fundamentals of biogas technology, fermenter designs, biogas purification, methanol production and utilization

Week 11: Organic Commodity Chemicals from Biomass: Biomass as feedstock for synthetic organic chemicals, lactic acid, polylactic acid, succinic acid, propionic acid, acetic acid, butyric acid, 1,3-propanediol, 2,3-butanedioil, PHA

Week 12: Integrated Biorefinery: Concept, corn/soybean/sugarcane biorefinery, lignocellulosic biorefinery, aquaculture and algal biorefinery, waste biorefinery, hybrid chemical and biological conversion processes, technoeconomic evaluation, life-cycle assessment

- Donald L. Klass, Biomass for Renewable Energy, Fuels, and Chemicals, Academic Press, Elsevier, 2006.
   Prabir Basu, Biomass Gasification, Pyrolysis and Torrefaction, Academic Press, Elsevier, 2013.
- 2. A.A. Vertes, N. Qureshi, H.P. Blaschek, H. Yukawa (Eds.), Biomass to Biofuels: Strategies for Global Industries, Wiley, 2010.
- 3. S. Yang, H.A. El-Enshasy, N. Thongchul (Eds.), Bioprocessing Technologies in Biorefinery for Sustainable Production of Fuels, Chemicals and Polymers, Wiley, 2013.
- 4. Shang-Tian Yang (Ed.), Bioprocessing for Value Added Products from Renewable Resources, Elsevier, 2007.

# Environmental Quality Monitoring & Analysis

#### **Course Duration: 12 weeks**

Credits: 3

## **Course layout**

Week 1: Introduction; Definition of Environment; Link between source/environment/receptor; Exposure; Health effects; Toxicology; Defining the need for fate and transport.

**Week 2**: Chemicals of concern; relevant properties for environmental fate and transport; Definition of Equilibrium – partition constants, solubility, vapor pressure, henry's constant, Koc, Kow etc. Equilibrium partitioning of chemicals between different phases of the environment.

Week 3: Parameters for environmental water/ air / soil / sediment – screening parameters, priority air pollutants – definitions of PM

**Week 4:** Monitoring of environmental parameters – screening parameters – BOD, COD, TOC, TDS; Environmental sampling – definition and synthesis of a monitoring/sampling/analysis method. Quality Assurance and quality control (QA/QC).

Week 5: Methods for sampling/processing/analysis of organic and inorganic constituents in air/water/soil/sediment.

Week 6: Introduction to environmental transport – BOX Models and the application to multimedia transport of pollutants

Week 7: Atmospheric Dispersion – Gaussian Dispersion model

Week 8: Fundamentals of mass transport – definition of intraphase and inter-phase chemical flux; interphase mass transport, diffusion coefficient and convenction mass transfer coefficients. Week 9: Chemical Exchange between air-water

Week 10: Chemical Exchange between sediment-water

Week 11: Chemical exchange between soil-air

Week 12: Overall transport model and scenarios

#### **Books and references :**

1. Environmental Chemodynamics - Louis J Thibodeaux, 2nd Edition, Wiley Interscience Environmental Engineering – Peavy, Rowe and Tchobanoglous, McGraw-Hill.

#### **Additional References:**

1. Atmospheric Chemistry and Physics – Seinfeld and Pandis.

# **Bio photonics**

#### **Course Duration: 12 weeks**

Credits: 3

## **Course layout**

Week 1: Introduction of Biophotonics

Week 2: Fundamentals of Light and Matter

- Week 3: Basics of Biology
- Week 4: Basics of light-matter interactions in molecules, cells and tissues
- Week 5: LASERs for Biophotonics
- Week 6: Bioimaging: Principles and Applications
- Week 7: Optical Biosensors
- Week 8: Light Activated Therapy: Photo Thermal and Photo Dynamic Therapy
- Week 9: Tissue Engineering with Light
- Week 10: Optical Tweezers, Scissors and Traps
- Week 11: Nanotechnology for Biophotonics: Nano Bio Photonics
- Week 12: Optogenetics & Neurophotonics

# Introduction to Environmental Engineering and Science – Fundamental and Sustainability Concepts

**Course Duration: 12 weeks** 

Credits: 3

**Course layout** 

Week 1: Sustainability Concepts – Innovations and Challenges

Week 2: Environmental Measurements from Different Disciplines

Week 3: Ecology, Population & Environmental Chemistry

Week 4: Physical Process in Environment

Week 5: Environmental Biological Concepts

Week 6: Environmental Risk Assessments with Concepts of EIA and LCA

Week 7: Water – Quantity and Quality

Week 8: Water Treatment Basics

Week 9: Basics of Wastewater Collection, Treatment & Resource Recovery

Week 10: Basics of Solid Waste, Soil and Noise Pollution

Week 11: Basics of Air Pollution Issues - Global and Local

Week 12: Case Studies and Course Wrap-up

- 1. Introduction to Environmental Engineering and Science by Gilbert M Masters and Wendell P Ela, Paperback: 696 pages, Publisher: Pearson Education India; 3rd edition, ISBN-10:9332549761, ISBN-13: 978-9332549760.
- 2. Environmental Engineering, by Howard Peavy, Donald Rowe, and George Tchobanoglous, Paperback: 736 pages, Publisher: McGraw Hill Education; ISBN-10: 9351340260, ISBN-13: 978- 9351340263

**Course Duration: 12 weeks Credits: 3** 

## **Course layout**

Week 1- Interactomics: Basics and Applications Lecture 1: Introduction to Proteomics Lecture 2: Introduction to Interactomics Lecture 3: High throughput platforms of interactomics: Protein arrays Lecture 4: Cell-free expression-based protein microarrays Lecture 5: NAPPA: Recombinational Cloning, Basic workflow, Surface Chemistry, Printing and Assessment

Week 2- Interactomics: Basics and Applications Lecture 6: NAPPA Technology and Protein Arrays-I Lecture 7: NAPPA Technology and Protein Arrays-II Lecture 8: Biomarkers: Harnessing the immune system for early detection of disease-I Lecture 9: Biomarkers: Harnessing the immune system for early detection of disease-II Lecture 10: Biomarkers: Harnessing the immune system for early detection of disease-III

Week 3- Interactomics: Basics and Applications Lecture11: NAPPA & its applications in study of antibody immune response in disease & in drug screening-I Lecture12: NAPPA & its applications in study of antibody immune response in disease & in drug screening-II Lecture13: NAPPA & its applications in study of antibody immune response in disease & in drug screening-III Lecture 14: Using functional proteomics to identify biomarkers and therapeutic targets-I Lecture 15: Using functional proteomics to identify biomarkers and therapeutic targets-II

Week 4- Interactomics: Basics and Applications Lecture 16: Applications of protein microarrays in Malaria Research-I Lecture 17: Applications of protein microarrays in Malaria Research-II Lecture 18: Introduction to Bioprinting and Iris<sup>™</sup> Optical QC Benefits-I Lecture 19: Introduction to Bioprinting and Iris<sup>™</sup> Optical QC Benefits-II Lecture 20: Screening of autoantibody signatures in cancer patients: Lab demonstration

Week 5- Interactomics: Basics and Applications

Lecture-21: Basics of Image Scanning and data acquisition Lecture-22: Applications of protein arrays in the identification of autoantibody signatures-I Lecture-23: Applications of protein arrays in the identification of autoantibody signatures-II Lecture-24: Applications of protein microarrays in deciphering PTMs and biological networks Lecture-25: Basics and Applications of Reverse Phase Protein Arrays-I

Week 6- Interactomics: Basics and Applications Lecture-26: Basics and Applications of Reverse Phase Protein Arrays-II Lecture-27: Basics and Applications of Reverse Phase Protein Arrays-III Lecture-28: An overview of label-free technologies Lecture-29: An overview of label-free technologies Lecture-30: Surface Plasmon Resonance- Principles and Assays-II Week 7- Interactomics: Basics and Applications

Lecture-31: Basics of SPR: Surface chemistry

Lecture-32: Basics of SPR: Experimental design

Lecture-33: Protein immobilization for protein-protein interaction studies

Lecture-34: Protein-protein interaction study: Binding analysis

Lecture-35: Protein-protein interaction study: Kinetic analysis

Week 8- Interactomics: Basics and Applications

Lecture-36: Use of SPR in unravelling domain motif interactions of proteasomal assembly chaperones

Lecture-37: Protein-small molecule interaction study: Immobilization & binding analysis

Lecture-38: Protein-small molecule interaction study: Kinetic analysis

Lecture-39: An introduction to biolayer interferometry (BLI) and its applications in protein research Lecture-40: Biomolecular interactions using Bio-Layer Interferometry (BLI)-I

Week 9- Interactomics: Basics and Applications

Lecture 41: Biomolecular interactions using Bio-Layer Interferometry (BLI)-II Lecture 42: Lab session- An introduction to Bio-Layer Interferometry (BLI) and its applications in protein research

Lecture 43: Applications of label-free technologies-II

Lecture 44: Biomolecular interaction analytics using Microscale

Thermophoresis Lecture 45: Mass Spectrometry coupled

Interactomics-I

Week 10- Interactomics: Basics and Applications

Lecture 46: Mass Spectrometry coupled Interactomics-II

Lecture 47: Next-Generation Sequencing Technology- Ion Torrent<sup>TM</sup>

Lecture 48: NGS Technology - Bioinformatics and data analysis-I

Lecture 49: NGS Technology - Bioinformatics and data analysis-II

Lecture 50: Next-Generation Sequencing Technology- Illumina

Week 11- Interactomics: Basics and Applications

Lecture 51: Agilent complete NGS target enrichment workflow for exomes, targeted panels and beyond

Lecture 52: The Human Pathology Atlas: A Pathology Atlas of the Human

Transcriptome-I Lecture 53: The Human Pathology Atlas: A Pathology Atlas of the Human Transcriptome-II Lecture 54: Statistical Analysis-I

Lecture 55: Statistical Analysis-II

Week 12- Interactomics: Basics and Applications

Lecture 56: Secondary Data Analysis

Lecture 57: Pathway Enrichment and Network Analysis

Lecture 58: Data Repositories and Databases

Lecture 59: Application of multi-omics approach for better understanding

of cancers Lecture 60: Integrated Omics and Systems Biology- Conclusion

#### **Books and references:**

Proteomics: A Cold Spring Harbor Laboratory Course Manual, A.J. Link and J. LaBaer, Cold Spring Harbor Laboratory Press, 2009. Selected papers from scientific journals Instructor bio

#### **Course Duration: 8 weeks**

Credits: 2

## **Course layout**

Week 1: Basic concepts of probability and statistics

Week 2: Basic concepts of linear algebra

Week 3: Basics of R

Week 4: Data visualization

Week 5: Correlation and regression

Week 6: Clustering and classification, Correlation and regression

Week 7: Clustering and classification

Week 8: Analysis of higher-dimensional data

#### **Books and references :**

1. Reading materials, links for online resources, Excel files and R codes will be provided by the instructor and will be adequate enough for this course.

#### **Reference books:**

- 1. Whitlock, Michael C.; Schluter, Dolph. The Analysis of Biological Data (2nd edition). Freeman, W. H. & Company, 2014.
- 2. Yang, Zheng R.; Machine Learning Approaches to Bioinformatics. World Scientific, 2010.
- 3. Moses, Alan; Statistical Modeling and Machine Learning for Molecular Biology. Chapman and Hall/CRC, 2016.
- 4. Hartvigsen, Gregg. A Primer in Biological Data Analysis and Visualization Using R, (1st Edition). . Columbia University Press, 2014.
- 5. Stewart, James; Day, Troy; Biocalculus: Calculus for Life Sciences. Cengage Learning, 2015
- 6. James, Gareth, etal. An introduction to statistical learning with application in R. Vol. 112. New York: springer, 2013.

First edition can be downloaded from the website https://www.statlearning.com/

## **Course Duration: 12 weeks**

## Credits: 3

# PREREQUISITES: This is an introductory course so anyone should follow it, basic knowledge of biology might be helpful

# **Course layout**

Week 1: Introduction: Flow of the history of biological inventions, basic Biological Macromolecules of life, i.e., Protein, Nucleic Acid, Carbohydrates & Lipid/Fat, and a comparison between polymers and "3C"ecrets of covalent bond, nucleic acid, DNA sequencing, PCR innovation, gene sequencing to genome sequencing, introduction to NGS and its different platforms, arrival of Post Genomic Era, the effect of HGP, and experimental three-dimensional structure determination techniques.

Week 2: Protein: Amino acids and their properties, Protein Chemistry, Chirality, Peptide bond, and Levels of protein structures, Dihedral angles, Peptide bond, and Ramachandran Plot, Super Secondary Structures, Motif, Domains, Non-covalent interactions, Folding of Protein, Thermodynamics, and Kinetics of protein folding, Characterization of Proteins.

**Week 3:** Introduction to Structural Biology Techniques: cellular organization, resolution structure determining technique with their ranges of the resolution, the success of X-ray crystallography from single molecule to a crystal, X-ray Crystallography, Crystallization in X-ray Crystallography, Crystallography.

Week 4: X-ray Crystallography: Production of X-ray and its properties, unit cell, symmetry, and lattice, the geometry of the crystal system, Crystal Symmetry, Instrumentation in X-ray Crystallography, Data collection, and processing

**Week 5:** X-ray Crystallography: Data Analysis of X-ray Crystallography - Diffraction Patterns, Indexing, Bragg's Law, Laue equation, Relation between "Laue equation and Bragg's Law", Lattice Transformation, Ewald Sphere, Laue Condition for Diffraction and Ewald Sphere, Structure Factors and Diffraction Pattern, Atomic Scattering Factor, Anomalous Dispersion, Analytical expression of the phase, Fourier Transformation, introduction to Phase Problem. Phase problem - Phase Problem, Patterson Function, How to solve phase problem, Heavy atom replacement methods, Isomorphous replacement, Anomalous dispersion, phase problem associated with crystal diffraction and common techniques to recover phase resolving different phase improvement methods. Refinement and Structure deposition to PDB - aspects of structure refinement, motivation, application, the procedure of simulated annealing, PDB repository, atomic model deposition as well as different PDB validation suites.

Week 6: NMR: Introduction to NMR, basic Principles of NMR and Instrumentation, NMR Sample Preparation and Chemical Shift related concepts, Factors effecting NMR Spectra (1D & 2D), 2D & 3D NMR Spectroscopy focusing on protein structure.

Week 7: Spectroscopy: Introduction to Spectroscopy, UV-Vis and CD spectroscopy, Fluorescence Spectroscopy and Green Fluorescence Protein (GFP), Infrared & Raman Spectroscopy for protein, Raman Spectroscopy, Raman Microscopy and Raman Crystallography for studying protein.

**Week 8:** Microscopy: Introduction to Microscopy, Functioning details of Cryo-Electron Microscopy (Cryo-EM), Cryo-Electron Microscopy: Data Collection and Analysis, A concise story of advancement Cryo-EM, Protein Data Bank.

Week 9: Molecular Visualizations: History of Molecular Visualizations of Biological Macromolecules, Description of structure-related files (.pdb, .mmcif, .mtz, etc.), Demonstration of COOT, 3D visualization using Pymol, Demonstration of Pymol.

**Week 10:** Molecular Dynamic Simulation: Why we need MD Simulation, Molecular Dynamic Simulation Process, Build a realistic atomistic model of the system, the algorithm behind simulation process, Concept of Topology and Parameter files, Major components in a force field, the concept of solvation, solvent models, Periodic Boundary Condition, Concept of Central Simulation Box, Phase Space, Concept of Ensembles, Energy Minimization (EM), potential energy surface (PES), Determination of EM, types of EM methods and their algorithms, Steps in MD Simulation, Application of Molecular Dynamic Simulation.

Week 11: Protein Engineering: What, How & Which of Protein Engineering, How to make logical ProteinEngineering: Process of Rational design, a success story of Rational Protein designing: Focusing on De NovoProcess, Designing Protein by mimicking nature: Process of Directed Evolution, Achievement, Challenges, andFuturedirectioninthefieldofProteinEngineering.

Week 12: Structure-Based Drug Discovery: Introduction to Structure-Based Drug Discovery (SBDD), Rational Drug Discovery, Docking Based Virtual Screening: Progress, Challenges and Future perspective, What makes a small molecule an ideal drug: Developing in silico ADMETox Model, Structure-Based Drug Discovery: Case study and Conclusion

- 1. Carl Ivar Branden and John Tooze., "Introduction to Protein Structure" 2nd 2001 Edition,
- 2. Taylor and Francis Voet, D. and Voet, J. G., "Biochemistry" 3rd edition,
- John Wiley and Sons.Introduction to Protein Architecture: The Structural Biology of Proteins, 2001 Arthur M. Lesk, Oxford University Press; 1st edition
- 4. Lubert Stryer, Biochemistry, 4th Edition, WH Freeman & CoCreighton.
- 5. T.E., Proteins, Structure and Molecular Properties, 2nd Edition, 1993
- 6. W.H. Freeman and Co McPherson, A. "Introduction to Macromolecular Crystallography", 2nd 2009 edition,
- 7. Wiley-Blackwell.Drenth, J., "Principles of Protein X-Ray Crystallography", 3rd edition, 2007 Springer.
- 8. Rhodes, G., "Crystallography Made Crystal Clear", 3rd edition, Academic Press

# **Biointerface Engineering**

#### **Course Duration: 8 weeks**

Credits: 2

**PREREQUISITES** : Bachelor Degree in any Engineering discipline

## **Course layout**

Week 1:Intermolecular Forces

Week 2: Adhesion and Wetting Phenomena

Week 3: Characterization of interfaces

Week 4: Protein-surface interactions

Week 5: Protein Aggregation

Week 6:Cell-surface interactions

Week 7: Surface modification and characterization

Week 8: Surface modification and characterization

#### **Books and references:**

• J. N. Israelachvili, Intermolecular and Surface Forces, 3rd edition, Academic Press, 2011.

• Willem Norde, Colloids and Interfaces in Life Sciences and Bionanotechnology, 2nd edition, CRC Press, 2011.

• W. Adamson, and A. P. Gast, Physical Chemistry of Surfaces, John Wiley, New York, 1997.

#### Duration:4weeks Credits:1

**Prerequisites:** Must have studied Mathematics at 10+2 level. Have studied graduate-level Biochemistry and Molecular Biology. Knowledge of Computer Programming will be helpful but not a necessity.

## **Course layout**

#### Week 1:

- 1. L1: Introduction to mathematical modeling in biology
- 2. L2: How to start modeling?
- 3. L3: Basic concepts of modeling using ODEs: Modeling the spread of infectious disease
- 4. L4: Basic concepts of modeling using ODEs: Modeling population growth
- 5. L5: Numerical solution of ODE-based models I
- 6. L6: Numerical solution of ODE-based models II

## Week 2:

- 1. L1: Simulating ODE-based models: Introduction to JSim
- 2. L2: Simulating ODE-based models: Examples of simulation in JSim
- 3. L3: Steady state and stability analysis: Understanding steady state
- 4. L4: Steady state and stability analysis: Stability of steady states
- 5. L5: Phase plane analysis I
- 6. L6: Phase plane analysis II

#### Week 3:

- 1. L1: Concepts of bifurcation
- 2. L2: Bifurcation in Biological systems
- 3. L3: Modeling molecular processes in cell
- 4. L4: Modeling molecular processes-I: Ligand-receptor binding
- 5. L5: Modeling molecular processes-II: Enzymatic reaction
- 6. L6: Modeling molecular processes-III: Transcription and translation

## Week 4:

- 1. L1: Modeling a signal transduction circuit: Negative feedback
- 2. L2: Modeling a signal transduction circuit: Positive feedback
- 3. L3: Modeling a signal transduction circuit: Incoherent feedforward 4. L4:
- Modeling transcriptional circuits I
- 5. L5: Modeling transcriptional circuits II
- 6. L6: Online resources for mathematical modeling in biology

- 1. Mathematical Modeling in Systems Biology: An Introduction, Brian P. Ingalls, MIT Press, 2013.
- 2. Modeling the Dynamics of Life: Calculus and Probability for Life Scientists, Frederick R. Adler, Brooks/Cole, 2012.
- 3. Biocalculus: Calculus for Life Sciences, James Stewart, Troy Day, Cengage Learning, 2015.

# **Conservation Geography**

## Duration: 12 weeks Credits: 2

**Prerequisites: Nil** 

**Course layout** 

Week 1: Introduction to Conservation Geography

Week 2: The Earth

Week 3: Lithosphere and landforms

Week 4: Atmosphere

Week 5: Hydrosphere

Week 6: Physical Geography in the Indian context

Week 7: Biosphere

Week 8: Conservation of biodiversity

Week 9: Human population and conservation

Week 10: Resources and conservation

Week 11: Economic Geography and conservation

Week 12:

Special topics in Geography and conservation

- 1. Modern Physical Geography, Strahler & Strahler
- 2. General Climatology, Howard Critchfield
- 3. Principles of Geomorphology, William Thornbury
- 4. RS-GIS with free software, Ankur Awadhiya Earth's Climate: Past and Future, William F. Ruddiman

Duration: 12 weeks Credits: 3

Prerequisites: Has cleared 10+2 with science

**Course layout** 

Week 1: Introduction

Week 2: Ecological structure

Week 3: Ecological interactions

Week 4: Ecological energetics

Week 5: Population Ecology

Week 6: Community Ecology

Week 7: Distribution & abundance

Week 8: Management of threatened species

Week 9: Human Ecology

Week 10: Ecology of change

Week 11: Applied Ecology

Week 12: Revision

#### **Books and references:**

1. Krebs, C. J. The experimental analysis of distribution and abundance. Ecology. New York: Harper and Row.

2. Odum, E. P., & Barrett, G. W. Fundamentals of Ecology. Philadelphia: Saunders

3. Selected articles / papers as referred to in the lectures.

## **Organic Chemistry in Biology and Drug Development**

## Duration: 12 weeks Credits: 3

## Prerequisites: Basic Organic Chemistry

## **Course layout**

Week 1: Introduction, Amino Acids, Peptides and proteins

Week 2: Peptides and proteins (contd.)

Week 3: Peptides and proteins (contd.)

Week 4: Proteins as biological catalyst, Concept of inhibition

Week 5: Nucleic acids,

Week 6: Metabolism, Synthetic biology, Molecular Biology

Week 7: Chemistry of cofactors/coenzymes

Week 8: Principle of drug design, Modern day drug discovery

Week 9: Chemistry of diseases and Drug development

Week 10: Chemistry of diseases and Drug development (contd.)

Week 11: Proton pump inhibitors, Gene replacement and delivery

Week 12: Revision and Problem solving

- 2. Biochemistry by Voet and Voet
- 2. Drug Design by R. Silverman
- 3. Medicinal Chemistry by G. L. Patrick

## Legal and Regulatory Issues in Biotechnology

Duration: 4 weeks Credits: 1

# Prerequisites: Bachelor degree students Prerequisite: anyone who is interested to know the legal aspects of biotechnology industry

**Course layout** 

Week-1: Regulation of Biotechnology Research

Week-2: Intellectual Property Rights and Life sciences (Agriculture, Pharma, Biotech)

Week-3: Biotech Product commercialization: Regulatory Approval Process

Week-4: Understanding technology transfer in biotech sector

- 1. Bucknell Duncan (ed.), I Pharmaceutical, Biotechnology and Chemical Inventions (Oxford University Press, 2011).
- 2. Cook M. Trevor, Pharmaceutical Biotechnology and the Law (Lexis Nexis, 2d ed. 2009).
- Cook M.Trevor, The Protection Of Regulatory Data In Pharmaceutical And Other Sectors (Sweet and Maxwell, 2000).
- Hardcastel Rohan, Law and The Human Body; Property Rights, Ownership and Control (Hart Publishing, 2007).
- 5. Valverde J.L. (ed.), Key Issues in Pharmaceutical Law (IOS Press, Vol. 9 2009).
- Drexl Josef, Nari Lee (ed.), Pharmaceutical Innovation, Competition and Patent Law; A Trilateral Perspective (Edward Elgar, 2013).
- 7. Verkey Elizabeth, Law of Plant Varieties Protection, 30-32 (Eastern Book Company, 1st ed. 2007).
- 8. Herring Jonathan, Medical Law & Ethics (Oxford University Press, 5th Ed., 2014).
- 9. Ventose Eddy, Medical Patent law- The Challenges of Medical Treatment (Edward Elgar, 2011).
- Krattiger Anatole, Mahoney T. Richard, et.al., II Intellectual Property Management in Health and Agricultural Innovation; A handbook of best practices (MIHR, Oxford Center for Innovation, 2007).
- 11. Emily Jackson, Medical Law, text, cases and Materials , (Oxford University Press, 4th ed. 2013)
- 12. Holy F Lynch, Effy Vayena and Urs Gasser, Big data, Health Law and Bioethics, Edited by I. G. Cohen, (Cambridge University Press, 2018).

## Nanomaterials and their Properties

## Duration: 12 weeks Credits: 3

Prerequisites: Knowledge of thermodynamics and atomic structure

**Course layout** 

Week 1: Introduction, Overview of nanostructures nanomaterials

Week 2: Multiscale hierarchical nanostructures

Week 3: Thermodynamics of Nanomaterials

Week 4: Thermodynamics of Nanomaterials

Week 5: Surfaces and interfaces in nanostructures

Week 6: Surfaces and interfaces in nanostructures

Week 7: Properties of nanomaterials

Week 8: Properties of nanomaterials

Week 9: Properties of nanomaterials

Week 10: Properties of nanomaterials

Week 11: Properties of nanomaterials

Week 12: Properties of nanomaterials

#### **Books and references**

1. M. F. Ashby, P.J. Ferreira, D.L. Schodek, Nanomaterials Nanotechnologies and Design, Butterworth-Heinemann.

3. Dieter Vollath, Nanomaterials: An Introduction to Synthesis, Properties and Applications, Wiley-VCH.

## **Ecology and Environment**

Duration: 8 weeks Credits: 2

**Prerequisites:** Nil

**Course layout** 

Week 1: Dr. B.S. Murty -Introduction (1), Sustainability Definition / Goals, Climate Change (2), Case Studies (3) (Eg: Dams, Chemicals, e-waste, IOT, Landfill siting etc)

Week 2: Dr. Sudhir Chella Rajan-Sustainability and Economics (3), Sustainability and Ethics (3)

Week 3: Dr. Ligy Philip-(Water Quality/ Waste Management), Water Quality and Treatment (3), Waste Management and Treatment (3)

Week 4: Dr. B. S. Murty (Water Management/ Resources), Urban Drainage, Water Resource Management, Impact of Climate Change

Week 5: Dr. Srinivas Jayanti (Energy)-Energy Demand / Resources (1), Pollution from Energy generation (1), Energy and Climate Change (Global Warming) (1), Energy and Sustainability (1), Long Range and Short Range Solutions (1) (Global vs. India)

Week 6: Dr. R. Ravi Krishna-Risk Assessment Definition (1), Pollutant Pathways / Safety/ Exposure (1), Liability /Examples (1), Life Cycle Assessment (2), Environmental Management and LCA (1)

Week 7: Dr. Sudhir Chella Rajan-Urban Planning / Sprawl (1), Challenges in Urban Planning, Transport (1), Energy (Smart Grid) (1), Waste (1), Governance (1)

Week 8: Dr. Susy Varughese / Dr. Parag Ravindran-Ecology – definitions / Systems (1), Biodiversity (1), Examples of Historical Impact of economy on Ecology, Restoration / Ecological Engineering

Week 9: Dr. Ligy Philip / Dr. Ravi Krishna -Solid Waste Management, Hazardous Waste Management

**Books and references:** Wrap up Emphasis on Climate Change and Adaptation

# **Bioreactors**

Duration: 4 weeks Credits: 1

### Prerequisites: Ability to appreciate simple mathematical analysis

**Course layout** 

Week 1: Introduction

Week 2: Two important outcomes of a bioprocess: biomass (cells) and bio-products

Week 3: Common bioreactor operation modes, factors that affect bioreactor performance

Week 4: The cell view of a bioreactor

### **Books and references:**

Shuler, M.L. and Kargi, F. 2002. Bioprocess Engineering: Basic Concepts, Prentice Hall, Englewood Cliffs, NJ J. Bailey and D. Ollis, Biochemical Engineering Fundamentals; McGraw Hill, 1986.

1. In addition, the students would be directed to specific sources during the course – they will become available during the course on the course page.

### **Transport Phenomena in Biological Systems**

### Duration: 12 weeks Credits: 3

### Prerequisites: Undergraduate engineering mathematics

### **Course layout**

Week 1: Introduction; Mass conservation principle

Week 2: Mass flux

Week 3: Mass flux contd.; Review

Week 4: Momentum flux

Week 5: Momentum flux contd.

Week 6: Momentum flux contd.

Week 7: Momentum flux contd.; Review; Energy (heat) flux

Week 8: Energy (heat) flux contd; Review

Week 9: Charge flux; Review

Week 10: Fluxes under simultaneous, multiple driving forces

Week 11: Fluxes under simultaneous, multiple driving forces contd.

Week 12: Fluxes under simultaneous, multiple driving forces contd.; Review

### **Books and references**

### **Textbook:**

1. Suraish kumar GK. 2014. Continuum Analysis of Biological Systems: Conserved Quantities, Forces and Fluxes. Springer, Heidelberg (e-book available free through Springer Link if your Institution has access to it).

### **References:**

- 1. Truskey, GA, Yuan F, Katz DF. 2009. Transport Phenomena in Biological Systems. II ed. Prentice Hall, New Jersey.
- 2. Bird, RB, Stewart, WE, Lightfoot, EN. 2001. Transport Phenomena, II edition, John Wiley and Sons, New York.

# **Technologies for Clean and Renewable Energy Production**

Duration: 8 weeks Credits: 2

### **Prerequisites: Nil**

### **Course layout**

- Week 1: Introduction, characterization of coal and conventional routes for energy production from Coal.
- Week 2: Cleaner routes for energy production form coal

Week 3: Characterization of crude oil and conventional routes for crude oil utilization

Week 4: Cleaner routes for energy production form petroleum crude

Week 5: Cleaner energy production from gaseous fuels

Week 6: Solar and wind energy production

Week 7: Production of hydro and geothermal energy

Week 8: Energy production from biomass and wastes and energy conservation

- 1. Miller Bruce G., Coal Energy Systems, Elsevier Academic Press, Paris 2005.
- 2. Twidel, J. and Tony W., Renewable Energy Resources, Second Edition, Taylor & amp; Francis 2006.
- 3. Kreith F., Goswami D.Y., Energy Management and Conservation, CRC Press 2008.
- 4. Sukhatme S., J Nayak J., Solar Energy: Principles of thermal Collection and Storage, 3 rd Ed., Tata McGrow-Hill Pulishing Company Ltd. 2008.
- 5. Mondal P and Dalai A., Sustainable utilization of natural resources, CRC Press 2017.

# Principles and Practices of Process Equipment and Plant Design

Duration: 12 weeks Credits: 3

### Prerequisites: Mass Transfer, Heat Transfer, Fluid Mechanics, Process instrumentation

### **Course layout**

**Week-1**: Introduction to Plant Design (2); Introduction to Mass transfer Equipment (1); Phase Equilibrium (2)

Week-2: Distillation – Fractionation (4); Design Problem (1)

Week-3: Flash Distillation (1); Batch Distillation (3); Design Problem (1)

Week-4: Absorption (2); Adsorption (2); Design Problem (1)

Week-5: Liquid-Liquid Extraction - 3; Column Internals – 2 [Sieve (1), Valve (1)]

Week-6: Column Internals contd. - Bubble Cap (2); packed column (1); Design Problem (2)

Week-7: Heat Exchanger: Introduction (1); Double Pipe HE (2); S&T HE (2)

Week-8: S&T HE contd. (1); Design Problem (1+2); Heat Exchanger Network (1)

Week-9: Heat Exchanger Network (3); Design Problem (2)

Week-10: Plant hydraulics: Pumps (2) Compressors (2), Pipeline (1)

Week-11: Pressure Vessels (2); Design Problem (2); Process Utilities (1)

Week-12: Safety (2), Process Design Package (3)

- 1. Process Equipment and Plant Design Principles and Practices", Ray. Subhabrata and Das, Gargi; ISBN: 9780128148853; 1st Edn., May 2020, Elsevier Inc.
- 2. Smith BD. Design of equilibrium stage processes. McGraw-Hill Companies; 1963.
- 3. Sinnott, R.K.and Towler, G., 2013. Chemical Engineering Design, Chemical Engineering Design.
- 4. Shah RK, Sekulic DP. Fundamentals of heat exchanger design. John Wiley & Sons; 2003 Aug 11.
- 5. Lestina, T. and Serth, R.W., 2007. Process heat transfer: Principles, applications and rules of thumb., Elsevier Ltd.

# **Introduction to Biomedical Imaging Systems**

# **Duration: 12 weeks**

### Credits: 3

Prerequisites: Engineering students from most core branch will be ready to take it in 6th-8th semester as they would have completed signals and systems and linear algebra.

### **Course layout**

Week 1: Introduction, 2D- Signals Systems review, Image Quality metrics

Week 2: Introduction, 2D- Signals Systems review, Image Quality metrics

Week 3: Projection Radiography

Week 4: Projection Radiography

Week 5:X-ray CT

Week 6: Nuclear Medicine- PET/SPECT

Week 7: Nuclear Medicine- PET/SPECT

Week 8: Ultrasound Imaging

Week 9: Ultrasound Imaging

Week 10: MRI

Week 11: MRI

Week 12: MRI

### **Books and references:**

1. Medical Imaging Signals and Systems by J. L. Prince and J. M. Links, Pearson Prentice Hall, 2006, ISBN 0130653535.

2. Webb's Physics of Medical Imaging, 2nd Edition, CRC press

# **Biomechanics of Joints and Orthopaedic Implants**

### **Duration: 8 weeks**

### Credits: 2

### Prerequisites: Engineering Mechanics, Solid Mechanics

### **Course layout**

**Week 1:** Introduction Musculoskeletal system Bone, Muscle, Ligament, Tendon, Cartilage and Meniscus – structure and function Anatomy of Synovial Joints – Hip, Knee, Shoulder, Elbow

Week 2: Biomechanics of Human Joints: (a) Hip Joint; (b) Knee Joint; (c) Shoulder Joint; (d) Elbow Joint

**Week 3:** Biomechanics of Gait cycle Gait Analysis Measurement techniques 3-D Motion analysis system – markers, cameras and force platform Lower extremity – hip musculoskeletal forces.

Week 4: Joint Kinematics Principle of Forward and Inverse Dynamics Calculations on joint forces and moments Calculations on muscle forces Model-based estimation of musculoskeletal forces during movements

**Week 5:** Concepts of Stresses and Strain Bone structure - Cancellous and Cortical Bone Mechanical Behaviour of Bone Bone Adaptation and Viscoelasticity Bone Anisotropy.

Week 6: Biomechanics of Joint Replacement – Hip, Knee, Shoulder, Spine Cemented and Cementless fixation Failure mechanisms of implants Implant Design Considerations

**Week 7:** Biomechanical modelling techniques and analysis Finite Element Analysis of bone and implant Bone Remodelling – formulation, algorithm, simulation Experimental validation of numerical models

**Week 8:** Bone Fracture Healing Tissue Differentiation Mechanoregulatory principle Mechanobiology based simulation of bone ingrowth around implants – acetabular and femoral components

- 1. Basic Biomechanics of the Musculoskeletal System" by Margareta Nordin and Victor H. Frankel
- 2. Biomechanics and Motor Control of Human Movement" by David A. Winter
- 3. Orthopaedic Biomechanics" by D.L. Bartel, D.T. Davy and T.M. Keaveny

# **Neuroscience of Human Movements**

### **Duration: 12 weeks**

Credits: 3

### Prerequisites: Motivation & open-mindedness is the only pre-requisite

# **Course layout**

- Week 1: Introduction, Membrane Physiology, Nernst Equation, GHK Equation, Action potential
- Week 2: Neuromuscular Junction, Skeletal Muscles
- Week 3: Skeletal muscles, Motor Units
- Week 4: Receptors, Muscle Spindles, Golgi Tendon Organs, Spinal control
- Week 5: Monosynaptic, Oligosynaptic & Polysynaptic reflexes
- Week 6: Preprogrammed reactions, Spinal control, Overview of motor control system, Primary Motor cortex – Part 1
- Week 7: Primary Motor cortex Part 2, Lesions, Brain Machine interfaces
- Week 8: Primary Motor Cortex Part 3, Role of Cerebellum in movement control
- Week 9: Role of Cerebellum in movement control
- Week 10: Parietal & Pre-motor cortex
- Week 11: Role of Basal Ganglia in movement control
- Week 12: Role of Basal Ganglia in movement control

#### **Books and references:**

1. Kandel & Schwartz, Principles of Neural Science, 2012, McGraw-Hill.

### **Duration: 8 weeks**

Credits: 2

**Prerequisites:** There is no pre-requisite to undertake this course. It is suitable for personnel with scientific/medical background (BSc/MSc/PhD/BPharm/MPharm/BAMS/BHMS/BDS/MDS/MBBS/MD/DM). Personnel working in the area of drug development/clinical trials/research may benefit from this course.

### **Course layout**

Week 1: Lecture 0: Course overview Lecture 1: Overview of Indian drug regulatory system Lecture 2: Overview of drugs & cosmetics Act and Rules thereunder Lecture 3: Overview of New Drug and Clinical Trials Rules Rules, 2019

Week 2: Lecture 4: Pre-clinical data requirements Lecture 5: Rules governing clinical trials Lecture 6A: Phases of clinical trial, forms, and fees Lecture 6B: Regulatory pathway and data requirements for NDCT, 2019

**Week 3:** Lecture 7: BA/BE study and study centres: Legal provisions Lecture 8: Guidelines to conduct BA/BE studies Lecture 9: Ethics Committee registration and re-registration

Week 4: Lecture 10: Ethical considerations Lecture 11: Good Clinical Practice Lecture 12A: Requirements for import/manufacture of new drug/IND for conducting clinical trials in India Lecture 12B: Requirements for import/manufacture of new drug/IND for sale/distribution and unapproved new drug for patients

Week 5: Lecture 13: Important issues Lecture 14: Special concerns Lecture 15: Clinical trial related guidelines (NDCT Rules)

Week 6: Lecture 16: Content of proposed clinical trial protocol Lecture 17: Content of a clinical trial report Lecture 18: Post marketing assessment and clinical trial compensation

Week 7: Lecture 19: Common observations during submission of CT/BA/BE protocol Lecture

20: Common observations during CT/BA/BE centre inspections Lecture 21: Drug development process: Overview

Week 8: Lecture 22: Salient feature of NDCT 2019 (What's new in NDCT?) Lecture 23A: Online submission (SUGAM) Lecture 23B: Online submission (CTRI) Lecture 24: Tables given in NDCT 2019 and its content

### **Books and references:**

1. Drugs & Cosmetics Act, 1940 and Rules thereunder 1945, New Drugs and Clinical Trial Rules, 2019, Related Guidance documents available at CDSCO website.

# **Functional Genomics**

### Duration: 4 weeks Credits: 1 Prerequisites: Basic level of understanding in cell and molecular biology is expected

### **Course layout**

Week 1: [2.5 hrs; 4 lectures] Introduction to Functional Genomics: Pre- and post-genomic era; major advancements in genomic approaches; epigenetics and metagenomics; forward versus reverse genetics

Week 2: [2.5 hrs; 4 lectures] Genome Analyses - Part 1

Genome editing approaches and their applications; gene expression analyses and applications

Week 3: [3 hrs: 4 lectures and 2 tutorial sessions] Genome Analyses - Part 2 Methods for DNA/RNA sequencing, sequence analysis and their applications

Week 4: [2.5 hrs: 3 lectures and 2 laboratory sessions] *Comparative Genomics* Genomic insight into evolution; power of comparative genomic analysis

### **Books and references**

Mostly publically available literature. Will be shared with the participants during the launch of the course.

# Aspects of Biochemical Engineering

**Course Duration: 12 weeks** 

Credits: 3

### **PRE-REQUISITES:** Mathematics in 10+2

### **Course layout**

Week 1: Microbiology, Biochemistry and Bioproducts

Week 2: Stoichiometry and Thermodynamics of biochemical reactions

Week 3: Kinetics of homogeneous chemical reactions

Week 4: Different types of bioreactors and reactor analysis

Week 5: Kinetics of enzyme catalyzed reactions using free enzymes

Week 6: Kinetics of enzyme catalyzed reactions using immobilized enzymes

Week 7: Kinetics of substrate utilization, product formation and biomass production of microbial cells

Week 8: Kinetics of substrate utilization, product formation and biomass production of microbial cells

Week 9: Design and analysis of activated sludge process and anaerobic digester. Scale up of bioreactor

Week 10: Transport phenomenon in bioprocess

Week 11: Air and medium sterilization

Week 12: Operation and Process control, Downstream processing, Economic analysis of biochemical processes and summary & conclusion

- 1. Chemical Reaction Engineering, Octave Levenspiel
- 2. Biochemical Engineering Fundamentals by Bailey and Ollis
- 3. Bioprocess Engineering Principles by Doran
- 4. Bioprocess Engineering Basic Concepts by Shular and Kargi
- 5. Biochemical Engineering by Blanch and Clark
- 6. Biochemical Engineering by Aiba, Humphrey and Millis

# **RNA Biology**

### **Course Duration: 12 weeks**

Credits: 3

### PREREQUISITES: Minimum qualification is bachelors in Science. Post graduation in Science is desirable.

### **Course layout**

Week 1: Introduction to RNA Biology and RNA world (Lectures 1-6)

Week 2: RNA as enzymes: The Ribozymes (Lectures 7-9)

Week 3: RNA Transcription (Lectures 10-14)

Week 4: RNA Processing and Life cycle (Lectures 15-17)

Week 5: Alternative RNA processing and editing (Lectures 18-23)

Week 6: RNA splicing, export and stability (Lectures 24-27)

Week 7: snRNA, rRNA, miRNA, siRNA processing, export and function (Lectures 28-33)

Week 8: Mechanisms of RNA decay and Non coding RNAs (lectures 34-39)

Week 9: Dosage compensation and X-inactivation (Lectures 40-44)

Week 10: Dosage compensation, Xist and ncRNA in imprinting (lectures 45-51)

Week 11: Telomere, telomerase and impact on genomes (lectures 52-57)

Week 12: Epitranscriptome and protein synthesis (Lectures 58-62)

### **Books and references:**

1.John F. Atkins et al (ed.), RNA Worlds: From Life's Origins to Diversity in Gene Regulation CSHL press (2011).

2.Gunter Meister (ed.), RNA Biology: An Introduction Wiley press (2011).

- 3. David Elliott and Michael Ladomery (ed.), Molecular Biology of RNA Oxford University Press (2011).
- 4. James Darnell (ed.), RNA: Life's Indispensable Molecule CSHL press (2011).

# Biomechanics

# **Course Duration: 12 weeks**

Credits: 3

# PREREQUISITES: High school physics & mathematics. Kindly note that this course does NOT assume knowledge of Engg Mechanics and Strength of Materials

# **Course layout**

Week 1: Introductory Mechanics – Statics and Dynamics – Basic Principles.

Week 2: The human body as a biomechanical system – basic terminologies

Week 3: Kinematics of muscles and joints - free-body diagrams and equilibrium, forces and stresses in joints

Week 4: Biomechanical analysis of joints of upper limb - Shoulder, Elbow, wrist, hand and fingers

**Week 5:** Upper limb as a mechanical system – analysis of reaching as movement of a multi-link serial chain – forward kinematics, analysis of fingertip forces as a parallel manipulator

Week 6: Biomechanical analysis of joints – Spine, Hip, Knee, Ankle.

Week 7: Introduction to Postural stability and Gait analysis.

Week 8: Gait analysis in health and disease - basics.

Week 9: Mechanics of Hard Tissues - Definition of Stress and Strain, Deformation Mechanics, structure and mechanical properties of bone - cortical and cancellous bones, Wolff's law of bone remodeling;

Week 10: Soft Tissues - Structure, functions, material properties – tendon function, elasticity in a tendon, models of non-linear elasticity in a tendon – physiological and non-physiological regimes, Davis' law of soft tissue remodeling.

Week 11: Visco-elastic properties of soft tissues, Models of visco-elasticity: Maxwell & Voight models.

Week 12: Basic Biofluid mechanics - Flow properties of blood in the intact human cardiovascular system.

# **Books and references**

- 1.David A. Winter, Biomechanics and Motor Control of Human Movement .
- 2. Margareta Nordin and Victor H. Frankel, Basic Biomechanics of the Musculoskeletal System.

3. Francisco Valero-Cuevas, Fundamentals of Neuromechanics.

4. Susan Hall, Basic Biomechanics.

5. Irving Hermann, Physics of Human Body.

### **Course Duration: 12 weeks**

# Credits: 3

# **PREREQUISITES: Basic Biochemistry**

### **Course layout**

Week 1: Introduction to Enzymes Lecture 1: Introduction to Enzymes Lecture 2: Basics of Enzyme Lecture3: Enzyme Classification (Part-I) Lecture4:Enzyme Classification(Part-II) Lecture 5: Enzyme Nomenclature

Week 2: Structure of enzyme Lecture 6: Enzyme Structure (Part 1) Lecture 7: Enzyme Structure (Part 2) Lecture 8: Enzyme Structure (Part 3)

Week 3: Enzyme Production (Part 1) Lecture 9: Cloning of Enzyme (Part 1) Lecture 10: Cloning of Enzyme (Part 2) Lecture 11: Over-expression in Host

Week 4: Enzyme Production (Part 2) Lecture 12: Extraction of enzyme Lecture 13: Purification Strategies (Part 1) Lecture 14: Purification Strategies (Part 2)

Week 5: Enzyme Production (Part 3) Lecture 15: Purification Strategies (Part 3) Lecture 16: Purification Strategies (Part 4) Lecture 17: Enzyme Characterization Approaches

Week 6: Enzyme catalyzed Biochemical reactions Lecture 18: Enzyme Catalyzed reactions (Part 1: Carbohydrate Metabolism) Lecture 19: Enzyme Catalyzed reactions (Part 2: Lipid and Protein Metabolism) Lecture 20: Enzyme Catalyzed reactions (Part 3: Detoxification)

Week 7: Enzyme-Substrate interaction Lecture 21:Enzyme-Substrate interaction (Part 1: Spectroscopic approaches) Lecture 22:Enzyme-Substrate interaction (Part 2: Isothermal Calorimetry) Lecture 23: Enzyme-Substrate interaction (Part 3: Surface plasma resonance)

Week 8: Enzyme assay system and Kinetics Lecture 24: Enzyme assay system Lecture 25: Enzyme Kinetics (Part 1) Lecture 26: Enzyme Kinetics (Part 2) Week 9: Enzyme Inhibitor Designing Lecture 27: Inhibitor designing (Part 1: Traditional approach) Lecture 28: Inhibitor designing (Part 2: Modern approach) Lecture 29: Inhibitor designing (Part 3: Computational approaches)

Week 10: Enzyme Inhibition kinetics Lecture 30: Enzyme Inhibition kinetics (Part 1) Lecture 31:Enzyme Inhibition kinetics (Part 2) Lecture 32: Enzyme Inhibition kinetics (Part 3)

Week 11: Enzyme Applications (Part 1) Lecture 33: Enzymes in Industrial setup (Part 1) Lecture 34: Enzymes in Industrial setup (Part 2) Lecture 35: Enzymes in catalyzing chemical reactions

Week 12: Enzyme Applications (Part 2) Lecture 36: Enzymes in medical field Lecture 37: Enzymes in environment field Lecture 38: Enzymes in drug discovery

# **Books and references:**

### Text book:

1. Enzyme Kinetics: Behavior and analysis of rapid equilbraium and steady state enzyme systems. Irwin H SEGEL. ISBN: 978-0-471-30309-1.

2. Biochemistry. 5th edition, Berg JM, Tymoczko JL, Stryer L. publisher: W H Freeman

3. Biochemistry, 2nd edition Reginald Garrett and Charles Grisham

4. Cornish-Bowden, A., Fundamentals of Enzyme Kinetics (revised ed.), Portland Press (1995).

5. Nelson, David L. (David Lee), 1942-. Lehninger Principles of Biochemistry. New York : W.H. Freeman, 2005.

6. Voet, D., Voet, J. G., & Pratt, C. W. (2008). Fundamentals of biochemistry: Life at the molecular level. Hoboken, NJ: Wiley.

7. Tymoczko, Lubert Stryer, and LubertStryer. Biochemistry. New York: W.H. Freeman, 2002. Print.

# **Reference Book:**

1. Enzymes Biocatalysis: Principles and Applications. ISBN 978-1-4020-8361-7.

2. Enzyme : Catalysis, kinetics and mechanisms. N.S. Punekar. ISBN 978-981-13-0784-3.

3. Enzymes 2nd Edition Biochemistry, Biotechnology, Clinical Chemistry by T Palmer P L Bonner

4. Color Atlas of Biochemistry 2nd edition by Jan Koolman and Klaus-Heinrich Roehm

# **Metabolic Engineering**

### **Course Duration: 8 weeks**

Credits: 2

# **PREREQUISITES : Basics of Microbiology, Biochemistry, Genetics**

# **Course layout**

Week 1:Introduction to Metabolic Engineering, Basic concepts; Scopes and Applications; Metabolism overview \_1 (Cellular Transport processes, Fueling Reactions)

**Week 2:**Cellular Metabolism Overview\_2 (Biosynthetic reactions, Polymerization, Growth Energetics); Regulation of Metabolic Pathways

Week 3: Reconstruction of Genome-scale metabolic network

**Week 4:** Examples of pathway manipulations by metabolic engineering : Ethanol, Aminoacids, antibiotics, vitamines, biopolymers, etc.

**Week 5:** Examples of pathway manipulations by metabolic engineering : Improvements of cellular properties, Biodegradation,

Week 6: Metabolic Flux Analysis: Flux Balance Analysis (FBA), Flux Variability Analysis, Flux Map

**Week 7:** Experimental determination of Metabolic Fluxes: Isotope labeled substrate, Isotope mapping Mapping Matrix, Isotope Distribution Vector

Week 8: Application of metabolic Flux Analysis

### **Books and references:**

1. Metabolic Engineering, Principles and Methodologies; G N Stephanopoulos, A A Aristidou, J Nielsen

2. Advances in Biochemical Engineering/Biotechnology; Metabolic Engineering, Volume Editor: Jens Nielsen

3. Systems Metabolic Engineering, Methods and Protocols; H S Alper

4. Metabolic Pathway design, A Practical Guide; P Carbonell

# **Introduction to Professional Scientific Communication**

### **Course Duration: 4 weeks**

# Credits: 1

# PREREQUISITES: Basic level of understanding on concept and methodology in scientific research

# **Course layout**

Week 1: Introduction to Professional Scientific Communication, Discussion of creativity, research ideas and where to find them, Inductive reasoning versus deductive reasoning

Week 2: Hypothesis, reasoning and testing the hypothesis, Peer review process, Structure of a scientific report

Week 3: Structure of a Research article, Title, abstract, methods, results, and discussion

Week 4: Structure of a Research article contd., Schematic diagrams, figures, tables and flow charts – rationale and usage, Ethics in biomedical research, Different forms of writing: scientific report, proposal, and reviews, Presentations-thumb rules and good practice

### **Books and references:**

Mostly publically available literature. Will be shared with the participants during the launch of the course.

# **Bioengineering: An Interface with Biology and Medicine**

# **Course Duration: 8 weeks**

# Credits: 2

# **PREREQUISITES: NIL**

# **Course layout**

Week 1 : Why biology for engineers: Part-I, Why biology for engineers: Part-II, Life processes & Cell, Cell and its properties, Clinician's Perspective-I

Week 2 : DNA Tools-Gene cloning, DNA Tools-Gene cloning-II, DNA Tools & Biotechnology, DNA Tools & Biotechnology-II,

Week 3 : DNA Tools & Biotechnology-III, DNA Tools & Biotechnology-IV, DNA Tools & Biotechnology-V, DNA Tools & Biotechnology-VI, Clinician's Perspective-III

Week 4 : Genetics-I, Genetics-II, Genetics-III, Genetics-IV, Clinician's Perspective-IV

Week 5 : Chromosomal basis of inheritance, Linkage, chromosomal disorders, Classical Genetics experiments, Bacteria and Viruses, Clinician's Perspective-V

Week 6 : Cell cycle, Cell cycle disregulation & Cancer, Developmental Biology, Principles and application of Animal Cloning, Evolution & Bioinformatics

Week 7 : Amino acids & proteins, Proteins & Proteomics, Techniques to Study Protein & Proteome-I, Techniques to Study Protein & Proteome-II, Techniques to Study Protein & Proteome-III

Week 8 : Techniques to Study Protein & Proteome-IV, Protein Interactions & Microarrays, Protein interactions & Systems biology, Bioinformatics, Ethics in Research and Publications

**Books and references:** Campbell Biology

# Environmental Biotechnology (being offered as elective in our R20 curriculum)

**Course Duration: 12 weeks** 

Credits: 3

# PRE-REQUISITE : Microbiology, Biochemistry, Genetics/Genomics

**Course layout** 

Week 1: Environmental Biotechnology and Sustainability. Scope and applications of the subject. Basics of ecosystem structure and function

Week 2: Microbial Ecology and Environmental Biotechnology: Concepts and importance of microbial ecology in Environmental Biotechnology

**Week 3:** Microbiology of Environmental Engineering System: Microbial diversity, growth and decay. Stoichiometry of microbial energetics and kinetics.

Week 4: Resource Exploitation by Microorganisms: Functions of various microbial groups relevant to environmental systems, including waste treatment and resource recovery, implications in biogeochemistry.

Week 5: Methods in Microbial Ecology with relevant to Environmental Biotechnology: Culture dependent and - independent analyses of microbial communities; PCR based methods, Microarray, Environmental genomics

**Week 6:** Microbial Principles of Biodegradation, Bio detoxification and other processes relevant for Environmental Applications: Microbial engines, (metabolism), Requirements for biodegradation, acclimation, Common biotransformation mechanisms; Effect of organic contaminant structure on biodegradability; Cooperation between different microbial species for enhanced biodegradation; Applying biodegradation kinetics to fate and transport modeling

Week 7: Bioremediation Technologies: Concepts, methods and applications of natural attenuation and engineered bioremediation (e.g bioaugmentation and biostimulation)

Week 8: Microbial Interactions with Heavy Metals and Metalloids: Bioremediation, Biohydrometallurgy and other aspects of Environmental Biotechnology

**Week 9:** Aerobic and Anaerobic Degradation of Aliphatic and Aromatic Compounds. Microbial interaction with plastics, antibiotics and others emerging pollutants.

Week 10: Microbially Enhanced Phosphorus and Nitrogen Removal

Week 11: Microbially Enhanced Oil Recovery; Microbial role in Carbon Storage and Capture (sequestration, conversion to useful biopolymers, etc.).

Week 12: Case studies : Bioremediation, Carbon Storage and Capture, Bioenergy.

- 1. Environmental Biotechnology, Principles and Applications by Bruce E Rittman and Perry L McCarty, McGrawhill Higher education.
- 2. Environmental Biotechnology Edited by Hans-Joachim Jördening and J Winter, WILEY-VCH Verlag Gmbh & Co.
- 3. Bioremediation and Natural Attenuation by Pedro J J Alvarage and Walter A Illman, Wiley Interscience.
- 4. Environmental Biotechnology, Vol 10 Handbook of Environmental Engineering, Edited by L K Wang et al, Humana Press.

### **Industrial Biotechnology**

### (Being offered as elective in our R20 curriculum)

### **Course Duration: 12 weeks**

Credits: 3

### **PRE-REQUISITES:**

Knowledge in microbiology

**Biochemistry and mathematics in 10+2 level** 

### **Course layout**

- Week 1: Introduction, Microbes and enzymes of industrial importance
- Week 2: Different types of bioreactors and bioreactor design
- Week 3: Microbial growth, substrate degradation and product formation kinetics, Tutorial 1
- Week 4: Instrumentation, Sterilization of air, media and reactor
- Week 5: Upstream and Downstream processing
- Week 6: Production of Oxy Chemicals I: Tax and non-tax alcohol, Brewing industry, Tutorial 2
- Week 7: Production of Oxy Chemicals II: Wine making, Vinegar and citric acid production, Tutorial 3
- Week 8: Production of Oxy Chemicals III: Antibiotics: Penicillin; Streptomycin
- Week 9: High fructose corn syrup, Cheese making, and Single cell production
- Week 10: Vaccines production and Metal leaching
- Week 11: Bioenergy- Gaseous fuels: Biohydrogen, Biomethane and Microbial fuel cell;

Liquid fuels: Bioethanol, Biodiesel and Biobutanol

Week 12: Aerobic and Anaerobic wastewater treatment processes, Tutorial 4

- 1.Industrial Microbiology by Samuel Cate Prescott and Cecil Gordon Dunn
- 2. Biochemical Engineering Fundamentals by Bailey and Ollis
- 3.Bioprocess Engineering Principles by Doran
- 4.Bioprocess Engineering Basic Concepts by Shular and Kargi
- 5.Biochemical Engineering by Blanch and Clark
- 6.Biochemical Engineering by Aiba, Humphrey and Millis
- 7.A textbook of Industrial Microbiology by Wulf Crueger and Anneliese Cruegen

# **Introduction to Biostatistics**

# **Course Duration: 8 weeks**

# Credits: 2

# PRE-REQUISITES: Basic knowledge of 12th standard mathematics is sufficient.

# Course layout

| Week 1: Lecture 1. Introduction to the course  |
|--|
| Lecture 2. Data representation and plotting  |
| Lecture 3. Arithmetic mean   |
| Lecture 4. Geometric mean  |
| Lecture 5. Measure of Variability, Standard deviation                                      |
| Week 2: Lecture 6. SME, Z-Score, Box plot  |
| Lecture 8. Kurtosis, R programming   |
| Lecture 9. R programming   |
| Lecture 10. Correlation  |
| Week 3: Lecture 11. Correlation and Regression   |
| Lecture 12. Correlation and Regression Part-II   |
| Lecture 13. Interpolation and extrapolation  |
| Lecture 14. Nonlinear data fitting   |
| Lecture 15. Concept of Probability: introduction and basics                                |
| Week 4: Lecture 16. counting principle, Permutations, and Combinations                     |
| Lecture 17. Conditional probability  |
| Lecture 18. Conditional probability and Random variables                                   |
| Lecture 19. Random variables, Probability mass function, and Probability density function  |
| Lecture 20. Expectation, Variance and Covariance   |
| Week 5: Lecture 21. Expectation, Variance and Covariance Part-II                           |
| Lecture 22. Binomial random variables and Moment generating function                       |
| Lecture 23. Probability distribution: Poisson distribution and Uniform distribution Part-I |
| Lecture 24. Uniform distribution Part-II and Normal distribution Part-I                    |
| Lecture 25. Normal distribution Part-II and Exponential distribution                       |
| Week 6: Lecture 26. Sampling distributions and Central limit theorem Part-I                |
| Lecture 27. Sampling distributions and Central limit theorem Part-II                       |
| Lecture 28. Central limit theorem Part-III and Sampling distributions of sample mean       |
| Lecture 29. Central limit theorem - IV and Confidence intervals                            |
| Lecture 30. Confidence intervals Part- II  |
| Week 7: Lecture 31. Test of Hypothesis - 1   |
| Lecture 32. Test of Hypothesis - 2 (1 tailed and 2 tailed Test of Hypothesis, p-value)     |
| Lecture 33. Test of Hypothesis - 3 (1 tailed and 2 tailed Test of Hypothesis, p-value)     |
| Lecture 34. Test of Hypothesis - 4 (Type -1 and Type -2 error)                             |
| Lecture 35. T-test   |
| Week 8: Lecture 36. 1 tailed and 2 tailed T-distribution, Chi-square test                  |
| Lecture 37. ANOVA - 1  |
| Lecture 38. ANOVA - 2  |
| Lecture 39. ANOVA - 3  |
| Lecture 40. ANOVA for linear regression, Block Design                                      |
|  |

- Introduction to Probability & Statistics Medenhall, Beaver, Beaver 14th Edition
   Introduction to Probability and statistics for engineers and scientists, S M Ross, 3rd Edition

# **Medical Image Analysis**

### **Course Duration: 12 weeks**

### Credits: 3

**PRE-REQUISITES:** Knowledge of calculus, Linear Algebra, introductory optimization and introductory matrix computations

### **Course layout**

Week 1: Introduction to medical imaging

- Week 2: Basic image processing techniques
- Week 3: Image registration 1- Rigid models
- Week 4: Image registration 2- Non-Rigid models
- Week 5: Image registration 3- Application and demonstration
- Week 6: Image segmentation Statistical shape model
- Week 7: Image segmentation PDE based methods
- Week 8: Image segmentation application and demonstration
- Week 9: Computer Aided Diagnosis Case Study 1
- Week 10: Computer Aided Diagnosis Case Study 2
- Week 11: Deep Learning for Medical image analysis 3D Convolutional Neural Networks
- Week 12: Deep Learning for Medical image analysis Generative models for synthetic data

# **Introduction to Developmental Biology**

### (Being offered as elective in our R20 curriculum)

#### **Course Duration: 12 weeks**

# Credits: 3

### PREREQUISITES: Knowledge of basic biology, molecular biology and genetics will be essential.

**Course layout** 

- Week 1: Developmental Anatomy life cycle; comparative and evolutionary embryology; fate mapping
- Week 2: Differential gene expression
- Week 3: Differential gene expression; Basic concepts of genetics
- Week 4: The concept of model organisms; Core genetic techniques
- Week 5: Cell-Cell communication in Development basic concepts of morphogenesis and cell signaling
- Week 6: Cell-Cell communication in Development the signaling pathways
- Week 7: Axis specification during Drosophila embryogenesis
- Week 8: Axis specification during Drosophila embryogenesis
- Week 9: Plant Development
- Week 10: Early mammalian development Cleavage and gastrulation
- Week 11: Early mammalian development Axis formation
- Week 12: Developmental mechanisms of evolutionary change

### **Books and references:**

Developmental Biology (9th or later editions) Author: Scott Gilbert

# **Tissue Engineering**

# (Being offered as elective in our R20 curriculum)

### **Course Duration: 8 weeks**

Credits: 2

PREREQUISITES : Understanding of basic cell biology would be helpful

### **Course layout**

Week 1: Introduction to tissue engineering

Week 2: Scaffolds: extracellular matrix, natural and synthetic polymers

Week 3: Hydrogels, bioceramics, scaffold fabrication

Week 4: Material characterization

Week 5: Cell source, isolation, growth, differentiation

Week 6: Cell adhesion, migration, signaling, bioreactors and challenges in tissue engineering

Week 7: Host integration, bioethics, Applications: Skin tissue engineering

Week 8: Applications: Bone tissue engineering, Vascular tissue engineering, and Corneal tissue engineering

- 1. Bernhard O. Palsson, Sangeetha N. Bhatia, Tissue Engineering, 2004, Pearson
- Robert A Brown, Extreme Tissue Engineering: Concepts and Strategies for Tissue Fabrication, 2013, Wiley Blackwell
- 3. W Mark Saltzman, Tissue Engineering: Engineering Principles for the Design of Replacement Organs and Tissues, 2004, Oxford University Press
- 4. John P Fisher, Antonios G Mikos, Joseph D Bronzino, Tissue Engineering, 2006, CRC Press
- 5. Robert Lanza, Robert Langer, Joseph Vacanti, Principles of Tissue Engineering, Third Edition, 2007, Elsevier Academic Press

# **Organ Printing**

### **Course Duration: 8 weeks**

Credits: 2

**PRE-REQUISITE:** Knowledge on Biomaterials and Human Anatomy and Physiology.

### **Course layout**

Week 1: Introduction to Bioprinting; different types of bioprinting techniques and their advantages and disadvantages

Week 2: 3D tissue designing and 3D tissue/organ printing; various process parameters and their role in bioprinting

Week 3: Introduction to bioinks; biomaterials used for bioink development with their merits and demerits

Week 4: Critical parameters of bioink formulations for bioprinting, modulation of bioink properties to control different processing conditions

Week 5: 3D bioprinted in vitro, in vivo, and ex vivo research models and techniques; in vitro manipulation of cells and biomaterials with a bioprinter to engineer tissues for regenerative medicine or in vitro tissue/organ models

Week 6: In situ bioprinting and 4D bioprinting with examples from recent literature

Week 7: Biofabrication-based strategies from bench-to-bed to address specific clinical problems

Week 8: Next step in bioprinting (challenges and future direction); ethical issues related to bioprinting

- 1. Atala et al., Essentials of 3D Biofabrication and Translation. 1st edition, ISBN-13: 978-0128009727.
- 2. Zhang et al., 3D Bioprinting and Nanotechnology in Tissue Engineering and Regenerative Medicine. 1st edition, ISBN 9780128005477.
- 3. Forgacs et al., Biofabrication Micro- and Nano-fabrication, Printing, Patterning and Assemblies, 1st Edition, ISBN 9781455728527.
- 4. Derby B. Printing and prototyping of tissues and scaffolds. Science. 2012. 338:921-6.
- 5. Seliktar D. Designing cell-compatible hydrogels for biomedical applications. Science. 2012. 336:1124-8.
- 6. Murphy SV, Atala A. 3D bioprinting of tissues and organs. Nature Biotechnology. 2014. 32:773-85.
- 7. Pati F, Gantelius J, Svahn HA. 3D Bioprinting of Tissue/Organ Models. Angewandte Chemie International Edition. 2016.55:4650-65.

# **Genome Editing and Engineering**

**Course Duration: 12 weeks** 

Credits: 3

**PREREQUISITES:** 10+2 with Biology and Chemistry

**Course layout** 

Week 1:Introduction to genetics and genetic engineering

Week 2: Breakage and Repair Of Genomic DNA

Week 3: Recombination

Week 4: Targeted genetic modification

Week 5:Zinc Finger Nuclease (ZFN) Technology

Week 6: Transcription activator-like effector nuclease (TALEN) Technology

Week 7: Clustered regularly interspaced short palindromic repeats (CRISPR)/Cas9 technology

Week 8: Applications of genome editing in treating human diseases

Week 9:Genome engineered Disease modeling

Week 10: Engineered immune cells for cancer therapy

Week 11: Personalized therapy; Challenges: safety and specificity

Week 12: Ethical concerns: Germ line gene editing

- 1. Harber, J. E., Genome Stability: DNA Repair and Recombination, Garland Science, 2013.
- 2. Yamamoto, T., Targeted Genome Editing Using Site-Specific Nucleases, Springer, 2015.
- 3. Zlatanova, J. and Holde, K. van, Molecular Biology: Structure and Dynamics of Genomes and Proteomes. Garland Science, 2015.
- 4. Yamamoto, T.(Ed.), Targeted Genome Editing Using Site-Specific Nucleases: ZFNs, TALENs, and the CRISPR/Cas9 System, Springer 2015.
- 5. Barrangou , R. and Oost, J. van der, CRISPR-Cas Systems: RNA-mediated Adaptive Immunity in Bacteria and Archaea , Springer, 2013.
- 6. Addgene, CRISPR 101:A Desktop Resource, January 2016
- 7. Alberts , B. , Johnson , A., Lewis , J., Morgan, D., Raff, M., Roberts, K.and Walter, P., Molecular Biology of the Cell, 6<sup>th</sup> Edn., Garland Science, 2014.

# Next Generation Sequencing Technologies: Data Analysis and Applications

**Course Duration: 12 weeks** 

Credits: 3

PREREQUISITES: Basic knowledge of programming and statistics

### **Course layout**

Week 1: Next Generation Sequencing (NGS) Technologies

Week 2: NGS data formats and data quality check (QC)

Week 3: Hands-on tutorial 1 - NGS data and quality check

Week 4: Read Mapping Algorithms

Week 5: Read Mapping Tool Hands-on and SAM files

Week 6: Variant detection and CNV analysis

Week 7: RNA sequencing experiment and data processing

Week 8: Differential expression analysis and multiple hypothesis testing corrections

Week 9: Gene Ontology (GO) and pathway enrichment analysis

Week 10: Hands-on tutorial 2 - RNA-seq data processing and differential expression analysis

Week 11: Genome assembly algorithms

Week 12: Application of NGS in epigenomic studies

### **Books and references**

1. High-Throughput Next Generation Sequencing, Methods and Applications. (Springer). Editors: Kwon, Young Min, Ricke, Steven C. (Eds.)

2. Next Generation Sequencing, Methods and Protocols, 2018, Volume 1712, Steven R. Head, Phillip Ordoukhanian, Daniel R. Salomon (Eds), Humana Press. ISBN : 978-1-4939-7512-9

3. Next Generation Sequencing and Data Analysis 2021, Melanie Kappelmann-Fenzl, Springer.ISBN : 978-3-030-62489-7

# **Course Duration: 12 weeks**

Credits: 3

# PREREQUISITES: 1st year college Mathematics and Biology

### **Course layout**

### Week 1: Introduction to Neurons

- 1) Neuron structure
- 2) Networks of Neurons and Synapses
- 3) System of neural processing
- 4) Basic structures in the brain
- 5) Sensory Executive Behavior systems

# Week 2: Excitable Membranes and Neural Activity

- 1) Membrane Potential and All or None Spike
- 2) Patch Clamp Techniques, Membrane Potential
- 3) Ion Channels
- 4) Current Injection Synapses
- 5) Single neuron activity

# Week 3: Point models: Hodgkin Huxley Equations (HHE)

- 1) Point and Compartmental Models of Neurons
- 2) Hodgkin Huxley Equations I
- 3) Hodgkin Huxley Equations II
- 4) Reducing the HHE and Moris-Lecar Equations (MLE) 5) Properties of MLE

# Week 4: Analysis of Neural Models

- 1) Phase Plane Analysis I
- 2) Phase Plane Analysis II
- 3) Analyzing HHE
- 4) Bifurcations
- 5) Other Point Models

### Week 5: Spike Trains: Encoding and Decoding - I

- 1) Random Variables and Random Processes
- 2) Spike Train Statistics and Response Measure
- 3) Receptive fields and Models of Receptive Fields
- 4) The Spike Triggered Average (Coding)
- 5) Stimulus Reconstruction (Decoding)

### Week 6: Spike Trains: Encoding and Decoding - II

- 1) Nonlinear approaches: Basics of Information Theory
- 2) Maximally Informative Dimensions
- 3) Discrimination based approaches
- 4) Measuring Spike Train Distances
- 5) Statistical Methods in Discrimination

### Week 7: Spike Trains: Encoding and Decoding - III

1) Examples-I: Encoding/Decoding in Neural Systems

- 2) Examples-II: Encoding/Decoding in Neural Systems
- 3) Neural Population Based Encoding/Decoding I
- 4) Neural Population Based Encoding/Decoding II
- 5) Examples: Population Based Encoding/Decoding

### Week 8: Plasticity - I

- 1) Synaptic Transmission and Synaptic Strength
- 2) Ways of Modification of Synaptic Strength
- 3) Types of Plasticity
- 4) Short Term Plasticity I
- 5) Short Term Plasticity II

### Week 9: Plasticity - II

- 1) Implications of Short Term Plasticity
- 2) Long Term Plasticity I
- 3) Long Term Plasticity II
- 4) Modeling Long Term Plasticity
- 5) Computational Implications

# Week 10: Plasticity - III, Modeling Phenomena with Plasticity

- 1) Adaptation
- 2) Attention
- 3) Learning and Memory I
- 4) Learning and Memory II
- 5) Developmental Changes

### Week 11: Plasticity - IV, Modeling Phenomena with Plasticity

- 1) Conditioning and Reinforcement Learning
- 2) Reward Prediction (Error)
- 3) Decision Problems
- 4) Learning and Memory II
- 5) Developmental Changes

# Week 12: Theoretical Approaches and Current Research

- 1) Optimal Coding Principles I
- 2) Optimal Coding Principles II
- 3) Theoretical Approaches to Understanding Plasticity
- 4) Current Topics I
- 5) Current Topics II

- 1. Theoretical Neuroscience: Computational and Mathematical Modeling of Neural Systems,, Dayan and Abbott
- 2. Signal and Systems, Oppenheim and Wilsky
- 3. Information Theory and Coding, Cover and Thomas 4. Nonlinear Dynamics and Chaos, Strogatz
- 5. Methods in Neuronal Modeling, Editors: Koch and Segev
- 6. Ion Channels of Excitable Membranes, Hille
- 7. Principles of Neural Science, Kandel and Schwartz

# Neurobiology

# **Course Duration: 4 weeks**

# Credits: 1

**PREREQUISITES:** Basic Class 10th level knowledge of biology and mathematics is sufficient. No pre-requisite courses are required.

### **Course layout**

### 1: Introduction

- · History of Neuroscience
- · Evolutionary perspective
- · Methods and tools in neuroscience
- · Brain structures
- 2: Electrical activity in the brain
- · Conductance and capacitance
- Time constant and length constant
- · Equilibrium potential
- 3: Active conductances
- · Ion Channels,
- · Active conductances
- · Action potential

# 4: Synapses

- · Chemical synapses
- · Gap-junctions
- · Synaptic integration
- 5: Sensory systems
- Tuning curves
- · Perception
- · Olfactory system

# **Books and references:**

Principles of Neural Science, 5th edition (Eric Kandel & colleagues)

# **Course Duration: 12 weeks**

Credits: 3

PRE-REQUISITE: Preferable, but not essential, to have taken the NPTEL Sensors and Actuators

### **Course layout**

- Week 1: Introduction to the Nervous System and Basic Structure of the Nervous System
- Week 2: Evolutionary Lessons in Nervous System Function and Hierarchy of Neural Function from the Cell to Large Networks
- Week 3: Signal Transmission, Analog Signal Processing, and Digital Signal Processing in the Nervous System
- Week 4: The OS, Servomechanisms, and Control Systems in the Nervous System
- Week 5: Theories of Learning and Mechanisms of Learning
- Week 6: Biological Basis of Contemporary Neural Networks and Neural Substrates for Contemporary Neural Networks
- Week 7: Computational Neurobiology, Brain-Computer Interfaces: Neuromodulation and Recordings: Brain-Computer Interfaces (BCI) Devices and Systems, Introduction to BCI Devices for Neural Recording and Stimulation.
- Week 8: Introduction to Neuro-biopotentials: EEG, EMG and ECoG: Introduction to biopotentials, Data Acquisition, Signal Acquisition, Conditioning, and Processing.
- Week 9: Introduction to the development of BCI devices I
- Week 10: Introduction to the development of BCI devices II
- Week 11: Microdevices for Neural Stimulation and Recording: Flexible Devices for ECoG Recordings and Neural Stimulation, Microneedles for Measuring Local Field Potentials (LFPs), and Bioresorbable Devices for ECoG Recordings.
- Week 12: Demonstration of BCI Devices: Packaging, Implantation, and Recording: Packaging Techniques for BCI devices, Implantation of BCI devices, Interfacing with Read-out Electronics and Recording ECoG Signals.

# **Books and references:**

1. Fundamentals of Microfabrication by Madou Marc J.

2. Silicon VLSI Technology: Fundamentals, Practice, and Modeling by James D. Plummer, Michael Deal, and Peter D. Griffin.

- 3. Fundamentals of Semiconductor Fabrication by S M Sze
- 4. VLSI Fabrication Principles: Silicon and Gallium Arsenide by S K Gandhi
- 5. VLSI Technology by S M Sze
- 6. Fundamentals of Microelectronics by B Razavi
- 7. Franco, S., 2002. Design with operational amplifiers and analog integrated circuits. New York: McGraw-Hill.
- 8. Pallas-Areny, R. and Webster, J.G., 2012. Sensors and signal conditioning. John Wiley & Sons.
- 9. An Introduction to the Event-Related Potential Technique
- 10. The Oxford Handbook of Event-Related Potential Components
- 11. The Art of Electronics, Horowitz & Hill, 3rd Edition.
- 12. Principles of Neural Science, Fifth Edition. Eric R. Kandel, Edited, James H. Schwartz, Edited, Thomas M.
- Jessell, Edited, Steven A. Siegelbaum, Edited, A. J. Hudspeth, Edited, Sarah Mack, Art Editor Instructor bio

# **Classics in Neuroscience**

### **Course Duration: 8 weeks**

### Credits: 2

PREREQUISITES: Basic course in neuroscience can help

### **Course layout**

### Week 1:

Topic 1: Introduction: Why Study History? Why the 1940s and 1950s? Topic 2: Genes: Starting with DNA

# Week 2:

Topic 1: Signaling Molecules: The First Growth Factor Topic 2: Signaling Molecules: The First Neurotransmitters in the Brain

# Week 3:

Topic 1: Cell Biology and the Synapse Topic 2: Physiology: The Action Potential

# Week 4:

Topic 1: Physiology: Synaptic Potentials and Receptor Potentials Topic 2: Functional Organization of Neurons and Dendrites

# Week 5:

Topic 1: Neural Circuits: Spinal Cord, Retina, Invertebrate Systems Topic 2: Neural Circuits: Cortical Columns and Cortical Processing

# Week 6:

Topic 1: Neural Systems: The Neural Basis of Behavior Topic 2: Learning and Memory: Donald Hebb, Brenda Milner, and H. M.

# Week 7:

Topic 1: Neurology: Foundations of Brain Imaging Topic 2: Neurosurgery: From Cushing to Penfield

# Week 8:

Topic 1: Neuropsychiatry: The Breakthrough in Psychopharmacology Topic 2: Theoretical Neuroscience: The Brain as a Computer and the Computer as a Brain

### **Books and references:**

Creating Modern Neuroscience: The Revolutionary 1950s by Gordon M Shepherd

# Human Behaviour

**Course Duration: 8 weeks** 

Credits: 2

# **PREREQUISITES: NIL**

# **Course layout**

- Week 1 : Introduction to the science of human behavior
- Week 2 : Sensation & Perception-I
- Week 3 : Perception-II, Learning
- Week 4 : Memory and Language-I
- Week 5 : Language-II and Emotion
- Week 6 : Intelligence
- Week 7 : Personality
- Week 8 : Social influence and cognition

- 1. Attkinson and Hillgard, Psychology: An introduction, Cengage Press
- 2. Cacioppo, J, Discovering Psychology, Cengage Learning
- 3. Baron, R, Psychology, Pearson Press.

# Microsensors, Implantable Devices and Rodent Surgeries for Biomedical Applications

**Course Duration: 12 weeks** 

Credits: 3

**PREREQUISITES :** Preferable, but not essential, to have taken the NPTEL Sensors and Actuators.

**Course layout:** 

Week 1 : Logistics in rodent experiments: Personal Protective Equipment, Procurement and quarantine of laboratory animals, Husbandry and Handling of rodents, Breeding and Care of Rats and Mice.

Week 2 : Rodents for Neural experiments: Species of rodents used in neural experiments, Rodent identification, and selection for experiments

Week 3: Rodent Neural Sensors and Data acquisition systems, Epileptic Seizure Detection and Classification

Week 4 : Microbial and Health monitoring for chronic neural experiments.

Week 5 : Neural system models in rodents – Epilepsy, Stroke, Motor movements, Parkinsonian models and application of rodent neural experiments, Microfabrication for neural engineering, PVD, CVD, Lithography, Etching

Week 6 : Behavioral setups for neural experiments, Different Brain-Computer Interfaces, Micro Electrode Array

Week 7 : Rodent Micro Neurosurgery I: Rodent anesthesia techniques, Introduction, Neuroanatomy – Applied and Comparative (rodent /monkey/ human) Equipments, Sterility and sterilization, Craniotomy, Microneurosurgical instruments and techniques, Sensor implantation and head mounting techniques, Closure techniques and challenges, Implant care for chronic recordings

**Week 8 : Rodent Micro Neurosurgery II:** Spinal anatomy, Spinal cord sensors and implantation surgeries, Peripheral neural anatomy, Peripheral neural surgeries, Peripheral neural sensors, and implantation, Different neural experiments as an example, Recovery process, Biological Basis of Contemporary Neural Networks and Neural Substrates for Contemporary Neural Networks

**Week 9 : Rodent neuropathology:** Euthanasia and Brain harvesting techniques. Neuropathological processing of the harvested brain, Electronic System Development for neural experiments

Week 10 : Challenges in neural signals and synchronization of behaviors, Brain Stimulation: Introduction and Applications, Experimental Protocol for Neural Experiments

**Week 11 :** Neural signal processing and post-processing, EEGLAB, ERPLAB, Event-Related Potentials: Introduction and Applications, ERP Extraction, Time-Frequency Analysis, Signal Interpretation

Week 12: Decoding techniques and challenges, Recent Trends in Neural Engineering

- 1. Pallas-Areny, R. and Webster, J.G., 2012. Sensors and signal conditioning. John Wiley & Sons.
- 2. An Introduction to the Event-Related Potential Technique
- 3. The Oxford Handbook of Event-Related Potential Components
- 4. The Art of Electronics, Horowitz & Hill, 3rd Edition.

5. Principles of Neural Science, Fifth Edition. Eric R. Kandel, Edited, James H. Schwartz, Edited, Thomas M. Jessell, Edited, Steven A. Siegelbaum, Edited, A. J. Hudspeth, Edited, Sarah Mack, Art Editor
 6. Paxinos and Watson: The Rat Brain

## **Education for Sustainable Development**

## **Course Duration: 12 weeks**

Credits: 3

**Course layout** 

Week 1: 1: Introduction to ESD

a) Introduction to UNESCO 17 Sustainable Development Goals (SDG)

- Week 2 : b) SD Goal-4- Quality Education for all
  - c) Education for Sustainable Development (ESD)
- Week 3: 2: ESD & SDGs ESD for achieving SDG- 4.7
  - a)Sustainable lifestyle
  - b) Human rights
- Week 4 : c) Gender equality
  - d) Promotion of peace & non-violence
  - e) Global citizenship
- Week 5: f) Leveraging cultural diversity for SDGs
  - 3. ESD & Sustainability ESD for achieving SDG-4.4
  - a) Technical & vocational skills for employability
- Week 6 : b) 21st Century competencies for global & decent jobs c) Sustainable entrepreneurship
- Week 7 : d) Promoting good mental health & wellbeing

e) Inclusive education & social transformation

- Week 8:4: ESD & Social Transformation ESD for promotion of
  - a) Responsible consumption & production
  - b) Peace & justice in the society
- Week 9: c) Sustainable cities & communities
  - d) Sustainable health practices & social wellbeing
- Week 10: 5: ESD & Sustainable education Sustainable education & global partnership
  - a) Educational policy & curriculum
  - b) Pedagogical practices & ICT
- Week 11: c) Educational research & social benefits

# d) Educational ecosystem & management

# Week 12 : Guest Lecture by International Faculty

# **Books and references:**

- 1. Issues and trends in Education for Sustainable Development: UNESCO Publication
- 2. Digital Pedagogy for Building Peaceful & Sustainable Societies: Blue Dot Publication
- 3. https://www.mdpi.com/journal/sustainability/ special\_issues/Entrepreneurship\_Education

## **Advances in Omics**

## **Course Duration: 8 weeks**

Credits: 2

PREREQUISITES: Basics of biological molecules

## **Course layout**

Week 1: Introduction to genomics: Historical perspective with examples from Human genome project and Advent of NGS. Genomic assembly approaches.

Week 2: Detailed discussion of the principles of sequencing technologies and comparison of advantages and disadvantages. Applications and Challenges in the use of NGS technologies

Week 3: Omics data avalanche: 1000 genomes project, ENCODE project, EXAC, TCGA

**Week 4:** Importance of evolutionary viewpoint in genomics, Signatures of selection in primates (with prominent examples from Human studies), Whole-genome duplication, comparative and population genomics, tests of selection (codon based and site frequency-based tests).

**Week 5:** Introduction to transcriptomics, proteomics and Integration of Multi-Omic data. Other types of omic datasets resulting from high-throughput use of assays (Ex: Repli-Seq, Ribo-Seq, Tag-Seq)

Week 6: Omics databases organization and utility. NCBI, UCSC genome browser, Short Read Archive, Proteome Exchange, Peptide Atlas, KEGG

Week 7: Introduction to linux, use of command line interface, Tutorial on analysis of NGS data (Genomics, Transcriptomics)

Week 8: Course summary and Exam

## **Books and references:**

1. Selected Readings: Latest papers and review articles in each topic (from journals including, but not limited to, Nature, Curr. Opin. Chem. Biol., Cell, Nature Biotech., Science, Trends series). This will be provided in the class.

# **Statistics for Biomedical Engineers**

**Course Duration: 12 weeks** 

Credits: 3

PREREQUISITES: Preferably Masters/Senior Undergraduate students with engineering/mathematics background

Course layout Week 1: Introduction to Biostatistics, applications of biostatistics, discussion of few use cases.

Week 2: Introduction to statistics, Need for statistics, Role of probability, Discussion of descriptive statistics

**Week 3:** Discussion of Mean, Median and mode, Introduction to probability theory, probability distributions, Expectations, Population variance, sample statistics, Inferential statistics

Week 4: Central limit theorem, Confidence intervals, Introduction to Hypothesis testing, Elements of Hypothesis testing, Large sample test, p-values

**Week 5:** Small sample test, T-distribution, Type I error, Type II error, Power of test, Chi-Square distribution, Hypothesis test using variance, Contingency test, Test of Independence, Probability plots

**Week 6:** Hypothesis test for two independent population, paired T test, F-distribution, Detailed discussion on ANOVA, Derivation of Mean Squared Treatment and Mean Squared Error in ANOVA, Sample problems

Week 7: Joint distribution, Covariance & Correlation between random variables, Simple Linear Regression, R-squared statistic, Confidence intervals for regression parameters, Multiple Linear Regression, Adjusted R-Squared statistic

**Week 8:** Logistic Regression, logit function, Derivation of log-likelihood function, Revisit ANOVA using linear regression, Derivation of ANOVA equations, Sample problems

Week 9: Introduction to Blocking, Randomized Complete Block Design, Latin square design, Sample Problems

Week 10: Graeco-Latin Square design, Introduction to factorial design,  $2^2$  factorial design, Discussion on interactions

Week 11: 2<sup>3</sup> factorial design, Derivation of relevant equations, Sample problems

Week 12: 2-Way ANOVA, Use cases, Derivations, Sample problems

# **Books and references:**

1.Biostatistics: A foundation for analysis in the health sciences, Daniel, W. W. and C, L. Cross 9ed Wiley. 2013.

2. Biostatistics for the Biological and Health Sciences, Triola and Triola, Pearson Addison Wesley

3. Montgomery, Douglas C., and George C. Runger. *Applied statistics and probability for engineers*. John wiley & sons, 2020.

4. Montgomery, Douglas C. Design and analysis of experiments. John wiley & sons, 2017.

# **Course Duration: 12 weeks**

Credits: 3

PREREQUISITES: High school with science

## **Course layout**

# Week 1 : Challenges of biosecurity

- Agriculture Security
- Livestock production
- Human health
- Pandemics
- Bioterrorism

## Week 2 : Need for economic sensors to meet the emerging threats of biosecurity

- Case study for Filoviruses (Ebola, Marburg) sensor
- Case study for Bacillus anthracis (Anthrax) sensor
- Case study of botulism toxin
- Case study of insulin

## Week 3 : Design of electrochemical sensors

- Galvanic cells
- Nernst equation
- Equilibrium constant
- Cells as a chemical probe
- Concepts of  $E^0$  and  $E^{0'}$

## Week 4 : Electrode design and printing of electrodes

- Selection of electrode materials for rapid sensing
- Bio-inspired electrode materials

## Week 5 : Electrode design and printing of electrodes

- Processing of electrode materials
- Printing of electrodes
- Design challenges of electrode integration in protective technical textiles and defence gears

## Week 6 : Electrodes, potentiometry

- Reference electrode
- Indicator electrode

## Week 7 : Electrodes, potentiometry

- Junction potential
- Ion-selective electrode
- Solid-state chemical sensors (Fields effect transistors)

## Week 8 : Redox titrations in electrochemical sensors

- Redox titration curves
- Determination of endpoints
- The oxidation state of the analyte

## Week 9 : Electro-analytical methods

- Basic electrolysis
- Electro-gravimetric analysis

## Week 10 : Electro-analytical methods

- Coulometry
- Amperometry
- Voltammetry

# Week 11 : Designing human-on-a-chip platforms for toxin detection, neurocomputing, robotics, and rehabilitation

- Concepts of whole cell biosensors
- Neuro-technology
- Action potential shape analysis in toxin detection & role of voltage gating molecules
- Micro-electrode arrays
- Field effect transistors

# Week 12 : Designing of a biosecurity facility

- Infrastructure
- Instrumentation
- Sample handling, quarantine protocols, biosafety levels, personnel safety and security
- In-house fabrication, printing, and micro-machining facility
- Data analysis & storage

## **Books and references:**

1. *The Problems of Biological Weapons*, Milton Leitenberg, Swedish National Defence College, Department of Security and Strategic Studies, ISBN: 9789189683273, 9189683277 (2004)

2. *Biotechnology Research in an Age of Terrorism*, Committee on Research Standards and Practices to Prevent the Destructive Application of Biotechnology, National Academic Press, ISBN: 9780309089777, 0309089778 (2004)

3. Popular Reading: A Taste for Poison: Eleven Deadly Molecules and the Killers Who Used Them, Neil Bradbury, St. Martin's Press, ISBN-10: 1250270758 (2022)

4. Popular Reading: Hot Zone, Richard Preston, Anchor, ISBN: 0-385-47956-5 (1994)

5. Measurement, Instrumentation, and Sensors Handbook, Electromagnetic, Optical, Radiation, Chemical, and Biomedical Measurement, Volume 2, Compilers: Halit Eren, John G. Webster; CRC Press (2017)

6. *Principles of Instrumental Analysis*, Douglas A Skoog, James Holler, Stanley R Crouch, ISBN: 9781337468039, 1337468037 (2017)

7. Instrumentation, Franklyn W Kirk & Nicholas R Rimboi, American Technical Society, ISBN: 9780826934208, 082693420X (1966)

8. *Quantitative Chemical Analysis* (Edition 8), Daniel C. Harris, Freeman Palgrave Macmillan International Edition, ISBN- 13: 978-1-4292-1815-3 (2010)

9. A Textbook of Quantitative Inorganic Analysis including elementary Instrumental Analysis (Edition 3), Arthur I Vogel, The English Language Book Society & Longmans, Green & Co Ltd (1961)

10. The electrochemical detection of bioterrorism agents: a review of the detection, diagnostics, and implementation of sensors in biosafety programs for Class A bioweapons, Connor O Brien, Kathleen Varty, Anna Ignaszak, Microsystems & Nanoengineering, Volume 7, Article Number: 16 (2021)

11. Nanoelectronics & Nanosystems, From Transistors to Molecular & Quantum Devices, Karl Goes, Peter Glosekotter, Jan Dienstuhl, Springer, ISBN: 3-540-40443-0 (2004)

12. *Enabling Technologies for Cultured Neural Networks*, Volume 1, Editors: David A. Stenger, Thomas M. McKenna, ISBN: 9780126659702, 0126659702 (1994)

13. Nanobioelectronics - for Electronics, Biology, and Medicine, Editors: Andreas Offenhausser, Andreas Offenhäusser, Ross Rinaldi, Springer, ISBN: 9780387094595, 0387094598 (1979)

# Advanced Fluorescence Microscopy and Image Processing

## **Course Duration: 12 weeks**

## Credits: 3

**PREREQUISITES:** A Bachelor's degree in any area of life sciences (zoology, botany, biochemistry, biotechnology, microbiology, medical biotechnology, genetics, etc.) is preferred. However, final year B.Sc./B.Tech/B.E. students in life sciences/Biotechnology can also enroll.

## **Course layout**

## Week 1:

Lecture 1: Introduction to fluorescence microscopy and its applications Lecture2:Introduction to Microscope Optics

## Week 2:

Lecture 3: Design of a Fluorescence Microscope Lecture 4: Fluorescent proteins, organic dyes, and protein labeling strategies

## Week 3:

Lecture 5: 6D imaging, live cell imaging, time-lapse imaging, FRAP, FRET, FLIM Lecture 6: Illumination strategies (Epi, TIRF, HILO, light sheet, multi-photon)

## Week 4:

Lecture 7: Confocal Microscopy Lecture 8: Super-resolution microscopy (SIM, STED, STORM/PALM)

# Week 5:

Live demonstration of a fluorescence microscope, 6D imaging of live cells

# Week 6:

Lecture 9: Immunofluorescence (IF), Immunohistochemistry (IHC) Lecture 10: Fluorescence in-situ Hybridization (FISH), RNAFISH

# Week 7:

Lecture 11: Digital images and camera technologies for microscopy Lecture 12: CCD, EMCCD, sCMOS camera

# Week 8:

Lecture13: ImageJ/FIJI-based image processing and data analysis (workshop, basic operations of ImageJ/FIJI)

# Week 9:

Lecture14: ImageJ/FIJI-based image processing and data analysis (workshop, image segmentation)

## Week 10:

Lecture 15: ImageJ/FIJI-based image processing and data analysis (workshop, macros, intensity measurement, scale bar, time scale, montage preparation, colocalization)

# Week 11:

Lecture 16: Single-molecule imaging and tracking Lecture 17: Optical tweezers and traction force microscopy

# Week 12:

Lecture 18: Spatial Transcriptomics and Proteomics (RNAscope, MERFISH, CODEX) Lecture 19: High-content imaging

# **Books and references:**

1) Microscope Image Processing by Qiang Wu, Fatima Merchant and Kenneth Castleman, Academic Press, ISBN: 012372578X

2) Fundamentals of Light Microscopy and Electronic Imaging by Douglas Murphy and Michael Davidson, Second Edition, Wiley-Blackwell publisher, ISBN: 047169214X.

3) Introduction to optical microscopy by Jerome Mertz, Cambridge University Press, 2nd Edition, doi: 10.1017/9781108552660, ISBN: 9781108552660

4) Fluorescence Microscopy: From Principles to Biological Applications by Ulrich Kubitscheck, 2nd edition, John Wiley & Sons publisher, eBook ISBN 9783527687725

e-resource: https://www.microscopyu.com/

e-resource: https://www.ibiology.org/online-biology-courses/microscopy-series/

e-resource: http://zeiss-campus.magnet.fsu.edu/index.html

e-resource: https://www.leica-microsystems.com/science-lab/topics/basics-in-microscopy/

e-resource: https://www.olympus-lifescience.com/en/microscope-resource/

e-resource: https://imagej.nih.gov/ij/docs/examples/index.html

# **Course Duration: 12 weeks**

Credits: 3

**PREREQUISITES:** Basic Biology Knowledge such as courses in Molecular Biology, Microbiology, Biochemistry, Genetics, etc

# **Course layout**

# Week 1:

Day 1: Introduction to Computational genomics, Transcriptomics, Proteomics, Epigenomics, Metagenomics and their applications, The BIG data of biological sciences

Day 2: Organization of genetic information in prokaryotic and eukaryotic cell, genome maps, Eukaryotic genome structure, High-throughput technologies to translate this information into genomic data

Day 3: How genomic data is organized in public databases, Genomics web resources, Nucleic acid and protein sequence databases, gene expression databases, Metabolic and metabolomic databases. Examples: NCBI GenBank and Expasy, EBI, Ensembl, UCSC, KEGG

# Week 2:

Day 1: First, second generation sequencing technologies including Sanger and Illumina and their data output Day 2: Long read sequencing and linked read sequencing (Nanopore, PacBio, TELL-Seq)

Day 3: Sequence formats: FASTA, GenBank, EMBL, XML, Fastq, fast5, etc., genomic database versions and archives, NCBI SRA, bio-project, accessions, data retrieval using wget, FTP, FileZilla, and scripts provided by the database team for genomic analysis

# Week 3:

Day 1: Introduction to Linux, basic commands for file handling

Day 2: Running jobs on Linux, processing, installation of genomic packages

Day 3: Introduction to R, commonly used packages, applications in genomic analysis

# Week 4:

Day 1: Introduction to genomes and packages for genomic analysis such as EMBOSS; Specifications of workstations needed for genomic analysis, Introduction to High Performance Computing and servers, and their need in genomic analysis

Day 2 : Overview and concepts in genomic and transcriptomic analysis of an organism with examples and case studies

Day 3: Sample collection, DNA extraction and quantification, and species identification of the species to be sequenced. RNA extraction and transcriptome sequencing approaches

# Week 5:

Day 1: Methods to estimate the amount of sequencing coverage needed for genomic assembly, use of hybrid sequencing approaches for appropriate coverage and assembly

Day 2: Short and long reads, paired-end reads, quality filtering of sequence data, Genome complexity assessment, Jellyfish and GenomeScope for generating k-mer count histograms and calculating genomic heterozygosity

Day 3: Concept of genome assembly, contigs, scaffolds, complete genome, draft genome, chromosomal level assembly, Genome assembly algorithms such as De-Bruijn graph, Overlap layout consensus (OLC), Hybrid assembly

# Week 6:

Day 1: Introduction to common assembly tools ABySS, SOAPdeneno, Flye, Supernova

Day 2: 10X genomic linked-read sequencing, use of proc10xG set of python scripts to pre-process the 10x Genomics raw reads, removal of barcode sequences

Day 3: Nanopore long reads analysis: Guppy for base calling of raw reads, adaptor removal using Porechop, Genome assembly workflow using three different assemblers: wtdbg, SMARTdenovo, and Flye, parameters for assembly

# Week 7:

Day 1: de novo assembly using Supernova, parameters, usage of genomic and transcriptomic reads to increase assembly contiguity

Day 2: Merging assemblies to create hybrid assembly, gap closing of assembly and polishing, fixation of small indels, base errors, and local misassemblies, determining the quality of assembly using N50, BUSCO scores, coverage etc.,

Day 3: Chromosomal level assembly using Hi-C, concept of reference genome, finished genome, draft genome, case studies

# Week 8:

Day 1: Annotation of repeats in final genome assembly using RepeatMasker, Determining the simple and complex repeat content of a genome

Day 2: de novo transcriptome assembly, Determining the coding gene set using MAKER pipeline

Day 3: Prediction of tRNA, rRNA and miRNA in a genome, Identification of metabolic pathways by KEGG

# Week 9:

Day 1: Comprehensive functional annotation of predicted genes or protein sequences by homology-based alignment using Blast or Blat, COGs, Gene ontology based annotation, Interproscan, PROSITE, Pfam, prints, patterns, motifs and fingerprints

Day 2: Evolutionary analysis using homologs, paralogs and orthologs, Multiple signs of adaptation, gene family expansion and contraction

Day 3: Taxonomic classification, marker sequences such as 16S rDNA and ITS, taxonomic hierarchy, Phylogeny reconstruction using multiple sequence alignment, Distance based approaches such as Neighbour joining, Character based approaches such as Maximum parsimony, Maximum likelihood, RAxML

# Week 10:

Day 1: Epigenetics, ChIp-seq, transcriptome and microarrays for regulation of expression

Day 2: Single cell genomics, 10X Chromium linked-reads and Illumina sequencing, single cell gene expression Day 3: Application of multiomics approaches in human health and diseases such as cancer, diabetes, etc.

# Week 11:

Day 1: Prokaryotic genome sequencing and assembly approaches, draft and complete genomes, taxonomic identification

Day 2: Gene prediction approaches and common methods, annotation of a bacterial genome, t-RNA, rRNA, operon prediction

Day 3: Phylogenetic, metabolic and comparative analysis

# Week 12:

Day 1: Microbiome and Metagenome, Human, organismal and environmental microbiomes

Day 2: Sequencing and assembly of metagenomes, gene prediction, annotation, MAGs

Day 3: Taxonomic analysis using amplicon sequence variants, Statistical analysis

# **Books and references**

1. Bioinformatics: Sequence and Genome Analysis by David Mount

2. Computational Genome Analysis: An Introduction by Richard C. Deonier, Simon Tavare, Michael S. Waterman, Springer India

# **Microbial Biotechnology**

## **Course Duration: 12 weeks**

# Credits: 3

PREREQUISITES: 10+2 with Biology and Chemistry

## **Course layout**

Week 1: Introduction and principles of microbial biotechnology, Classification and taxonomy of microbes

Week 2: Structure and life cycle of representative groups of bacteria, viruses and eukaryotic microorganisms

Week 3: Physiology of microorganisms, adaptation to diverse environmental conditions

Week 4: Genome structure, mechanisms of gene expression and regulation

Week 5: Control of microorgansisms

Week 6: Industrial and pharmaceutical applications of microorganisms

Week 7: Microbes in agriculture, biofertilizer, microbial pesticides, integrated pest management

Week 8: Environmental biotechnology

Week 9: Food production involving microorganisms and their products

Week 10: Microbes in medical biotechnology

Week 11: Microbes in alternative energy

Week 12: Patenting in microbial biotechnology

## **Books and references**

1. Lee Y. K., Microbial Biotechnology: Principles and applications. World Scientific Publisher, 2013.

2. Tortora, Funke and Case, Microbiology, An Introduction, 5th Edition. Benjamin/Cummings Publishing Company, Redwood City, CA, 1995.

3. Board RG, Jones D, Skinner FA, Identification methods in applied and Environmental Microbiology, 1st Ed. Blackwell Science, 1992.

4. Funke, Study Guide for Microbiology, 5th Ed. Benjamin/Cummings Publishing Company, Redwood City, CA, 1995.

# Optical Spectroscopy and Microscopy: Fundamentals of Optical Measurements and Instrumentation

**Course Duration: 12 weeks** 

Credits: 3

# **Course layout**

**Week 1**: Essential Quantum Mechanics: Uncertainity Principle, Probabilistic nature of measurement, postulates of qmech, Stern Gerlach equivalent in light, Photon picture (PMT response), Linear Vector Space.

**Week 2:** Time dependent perturbation theory, Fermi Golden Rule, Transition probability in light matter interaction, Beer Lambert relation, Einestin's phenomenolgical treatment, A and B coefficients, Spontaneous emission, Origins of fluorescence

Week 3: Nature of Fluorescence, Emisson spectrum, Absorption spectrum, Anisotropy, Life time, FRET

**Week 4:** Second quantaisation, creation and anhilation operators, Fock states, light matter interaction in Feynman digrams

Week 5: Spontaneous emission orgin, Stimulated Emission origin dependence through Fock states

Week 6: Laser emission, two state, three state and four state laser systems

**Week** 7: Real world lasers, Charecteristics of laser emission, thershold behavior, Laser gain equation, CW operation, Pulsed lasers, Qswitching, mode locking, Saturable absorber

**Week 8:** Laser induced fluorescence, optical components (lenses, mirrors, gratings, prisms) and their working principles, Interference filters, dichroic filters, efficiency calculations for SNR improvement, aligning an optical equipment.

**Week 9:** Intro to optical hardware, common opto-mechanical assemblies, setting up a simple laser based spectrometer using gratings in lab, calibration and acquisition of fluorescein spectra.

**Week 10:** Principles of photo detection, QE, Dynamic range shot noise, photodetectors – PMTs, photodiodes, photo resistors, understanding common metrics and specs. Detection electronics – preamps, A2Ds

**Week 11:** Area detectors, CCDs, emCCDs, sCMOS, comparison, read noise, speed and other sensor charecteristics. Theory of Image formations – widefiled microscopy, bright field, phase contrast, DIC and fluorescence microscopy

Week 12: Scanning system: Principles of scanning system, Gaussian light progation and focussing, optical resolution, deinition in xy and z. Measurement and charecetrisation in lab. Scanning as time averaged focus, optical hinges, imaging of hinges, Confocal microscope

# **Books and references:**

1. Optical Spectroscopy (Demtroder), Quantum Electronics - Yariv, Building Scientific Apparatus - WJ Moore

# Biological Data Analysis and Visualization with R

**Course Duration: 8 weeks** 

Credits: 2

PREREQUISITES: Should be at least at the UG level with knowledge of probability and statistics

**Course layout** 

Week 1: Introduction and set up for biological data analysis with R

Week 2: Basic statistical analysis and data visualization techniques

Week 3: Bioconductor packages

Week 4: Gene expression analysis and co-expression network

Week 5: Analysis of ChIP-seq data in R

Week 6: Regression models on biological data

Week 7: Dimensionality reduction techniques

Week 8: Decision trees and Random Forest

## **Books and references:**

1. Introduction to Bioinformatics with R: A Practical Guide for Biologists (Chapman & Hall/CRC Computational Biology Series)

2. R Programming for Bioinformatics (Chapman & Hall/CRC Computer Science & Data Analysis) \

3. A Little Book of R for Bioinformatics 2.0 (brouwern.github.io)

# **Experimental Nanobiotechnology**

## **Course Duration: 4 weeks**

## Credits: 1

**PREREQUISITES:** Basic knowledge in biology / nanotechnology Or Desirable: NPTEL course on Biomedical nanotechnology

## **Course layout**

Week 1: Physical method of nanoparticle synthesis: Ball milling, Chemical synthesis of nanoparticles: Gold Nanoparticles, Hydrothermal synthesis of Carbon Dots, Polymeric nanoparticle synthesis (Albumin & PLGA Nanoparticles), Hydrogel fabrication

**Week 2:** Nanoparticle characterization techniques: UV-Vis Spectroscopy, DLS- Hydrodynamic Size & Zeta potential, Fourier transform infrared spectroscopy, Fluorescence Spectroscopy, Electron microscopic analysis

Week 3: Biomedical applications: Nanobiosensor, Fabrication of nanofibers using electrospinning, Hemocompatibility test, 3D Bioprinting, In vitro 3D cell culture.

Week 4: Nanotoxicology: Antibacterial properties of nanomaterials, In vitro cytotoxicity analysis, Apoptotic studies, In vivo toxicity studies using zebrafish embryo and hydra.

## **Books and references**

1.McNeil, S.E., (2011) Characterization of Nanoparticles Intended for Drug Delivery", Humana press

2.Xian, W. (2009). A laboratory course in biomaterials. CRC Press.

3. Micou, Melissa Kurtis, and Dawn Kilkenny. A Laboratory Course in Tissue Engineering. CRC Press, 2016.

4.Bisen, P.S., (2014) "Laboratory Protocols in Applied Life Sciences", Taylor & Francis Group, LLC

5.Holtzhauer, M., (2006) "Basic Methods for the Biochemical Lab", Springer-Verlag Berlin Heidelberg

# Algorithms in Computational Biology and Sequence Analysis

#### **Course Duration: 12 weeks**

## Credits: 3

**PRE-REQUISITES :** Elementary knowledge of discrete mathematics, basic algorithms and data structures is required. Programming proficiency with either C or C++ or Java or Python is required. Knowledge of basic algorithms for sorting, searching, hashing, graph traversal algorithms will be required.

## **COURSE PLAN :**

Week 1: Introduction

Week 2: Strings and exact matching

Week 3: Pairwise Sequence Alignment

Week 4: Heuristic Sequence Alignment

Week 5: Genome reconstruction using graph algorithms

Week 6: Evolutionary tree construction

Week 7: Sequence models and classification

Week 8: Pangenome graphs

Week 9: Discussion of research papers

Week 10: Discussion of research papers (Contd.)

Week 11: Discussion of research papers (Contd.)

Week 12: Discussion of research papers (Contd.)

## **Biophotonics**

## **Course Duration: 12 weeks**

Credits: 3

**PREREQUISITES :** None but basic knowledge of optical physics will be useful

## **Course layout**

- Week 1 : Introduction of Biophotonics
- Week 2 : Fundamentals of Light and Matter
- Week 3 : Basics of Biology
- Week 4 : Basics of light-matter interactions in molecules, cells and tissues
- Week 5 : LASERs for Biophotonics
- Week 6 : Bioimaging: Principles and Applications
- Week 7 : Optical Biosensors
- Week 8: Light Activated Therapy: Photo Thermal and Photo Dynamic Therapy
- Week 9 : Tissue Engineering with Light
- Week 10: Optical Tweezers, Scissors and Traps
- Week 11 : Nanotechnology for Biophotonics: Nano Bio Photonics
- Week 12 : Optogenetics & Neurophotonics

## **Books and references:**

- 1. Introduction to Biophotonics, Paras N. Prasad (2003)
- 2. Biophotonics: Concepts to Applications, Gerd Keiser (2016)
- 3. Physical Biology of the Cell, Rob Phillips (2012)
- 4. Fundamentals of Biomedical Optics, Caroline Boudoux (2017)

# **Comprehensive Molecular Diagnostics and Advanced Gene Expression Analysis**

#### **Course Duration: 12 weeks**

#### Credits: 3

## **Course layout**

**Week 1:** Basic Concepts including Central Dogma in Molecular Biology Definition and Scope of Molecular Diagnostics and Historical Developments Importance and advantages of molecular diagnostics over traditional methods, Nucleic Acid Structure and Function, DNA Replication and Repair, RNA Transcription and Processing, Protein Synthesis from mRNA-Translation

Week 2: Tools of Molecular diagnostics and Gene expression Analysis (I) PCR (Polymerase Chain Reaction) Fundamentals, RT PCR and qPCR, Modifications of PCR-Hot start, Touch down, nested PCR, Multiplex, Modifications of PCR 2-Long-range PCR,Single-cell PCR,Fast-cycling PCR, Methylation-specific PCR (MSP), Digital Droplet PCR-modern implications, PCR-based mutation analysis

Week 3: Tools of Molecular diagnostics and Gene expression Analysis (II) Principles (Sanger sequencing, Overview of NGS Technologies and Platform, Application of NGS in Molecular Diagnostics, Clinical Interpretation of NGS Data, Whole genome vs Whole exome sequencing, Targeted gene panels, NGS library preparations)

**Week 4:** Tools of Molecular diagnostics and Gene expression Analysis (III) DNA Microarray, FISH (Fluorescence in situ Hybridization), Serial analysis of gene expression, RNA sequencing, Tiling array, DNA protein interaction-chromatin immune precipitation.

**Week 5:** Techniques of Gene Manipulation: RNA interference and detection methods, Recombinant DNA Technology, CrispR-CAS9 technology, Epigenetics and diseases, DNA methylation analysis.

Week 6: Proteomics: Clinical Applications Overview of proteomics techniques and workflows, Protein separation techniques-brief discussion of gel electrophoresis and

chromatography, mass spectrometry, label-free and isotope labelling methods, role of metabolomics in laboratory diagnosis.

Week 7: Proteomics: Advanced topics in Clinical Proteomics High throughput proteomics like-Shotgun and data independent acquisition (DIA), Single cell proteomics and spatial profiling, methods to detect post translational modification and protein-protein interaction, proteomic data analysis and bioinformatic tools, Luminex multiplex assays and its application in biomarker analysis.

**Week 8:** Molecular Diagnostics in Infectious Diseases Syndromic Panels and Multiplex Assay, Molecular identification of Microorganism- covering bacterial, viral, fungal and parasitic diseases, antimicrobial resistance testing, POC Molecular diagnostics for infectious diseases, Molecular diagnostics in Hospital acquired infections.

**Week 9:** Molecular Diagnostics in Cancer Management Cancer markers, Liquid biopsies in cancer detection, circulating Tumour DNA (ctDNA) analysis, Monitoring treatment response with molecular diagnostics, Molecular diagnostics in targeted therapy, Digital PCR, Molecular diagnostics quality control

Week 10: Molecular Diagnostics in Genetic and Inherited Disorders Genetic testing and inherited diseases, Non-Invasive Prenatal testing (NIPT) and reproductive genetics, Molecular diagnostics in rare genetic disorders, Pharmacogenomics and Personalized Medicine, genetic counselling and patient education.

Week 11: Molecular Diagnostics in Medicine Molecular diagnostics in Metabolic disease, Molecular diagnostics in Neurodegenerative disease, Molecular diagnostics in Respiratory, Molecular diagnostics in Gastrointestinal

disorders, Molecular diagnostics in Endocrine disorders, Molecular diagnostics in Autoimmune disorders, Molecular diagnostics in Cardiovascular diseases, Molecular diagnostics in Transplantation diseases

Week 12: Molecular Diagnostics: Quality control and Ethical Concerns in and Futuristic Trends Quality control in molecular diagnostics, Ethical Concerns in Molecular Diagnostics, Microfluidics and Lab-on-chip in molecular diagnostics, AI and ML in molecular diagnostics, Nanotechnology based molecular diagnostics, Single cell Analysis, Integration of Multi-omics Data.

#### **Books and references:**

1. Molecular Cloning: A Laboratory Manual by David W. Russell and Joseph Sambrook

2. "Molecular Diagnostics: Fundamentals, Methods, & Clinical Applications" by Lela Buckingham and Maribeth L. Flaws

3."PCR (The Basics)" by Michael L. Mader

4. "Real-Time PCR: Advanced Technologies and Applications" by Nick A. Saunders and Martin A. Lee

5. "Next-Generation Sequencing: Translation to Clinical Diagnostics" by Alireza Heravi-Moussavi

6. "Genetic Testing and Molecular Biomarkers" by George P. Patrinos and William B. Coleman

7. "Essentials of Genomic and Personalized Medicine" by Geoffrey S. Ginsburg and Huntington F. Willard

8. "Bioinformatics for Beginners: Genes, Genomes, Molecular Evolution, Databases and Analytical Tools" by Supratim Choudhuri

9. "Genomic Medicine: Principles and Practice" by Dhavendra Kumar

10. "Pharmacogenomics: Challenges and Opportunities in Therapeutic Implementation" by Urs A. Meyer and FolefacAminkeng

11. "Cancer Genomics: From Bench to Personalized Medicine" by Graham Dellaire and Jason N. Berman

12. "Introduction to Genetic Analysis" by Anthony J.F. Griffiths, Susan R. Wessler, Sean B. Carroll, John Doebley

13. "Medical Genetics: A Core Text with Integrated Cases" by Lynn B. Jorde, John C. Carey, Michael J. Bamshad

14. "Molecular Diagnostics: Techniques and Applications for the Clinical Laboratory" by Wayne W. Grody and Robert M. Nakamura

15. "Molecular Pathology in Clinical Practice" by Debra G.B. Leonard

16. "Diagnostic Molecular Pathology: A Guide to Applied Molecular Testing" by William B. Coleman and Gregory J. Tsongalis

# **Cellular Biophysics: A Framework For Quantitative Biology**

**Course Duration: 8 weeks** 

Credits: 2

PRE-REQUISITES: BSc/BE/BTech 2nd year; BSc level knowledge of Classical mechanics; MSc level knowledge of Cell and molecular biology; MSc level knowledge of Biochemistry; Basic python programming

## **Course layout**

Week 1: Concepts in fluid dynamics as they apply to cellular scale life

Week 2: Diffusion & Macromolecular crowding

Week 3: Dynamics of macromolecules: Cytoskeleton

Week 4: Molecular motors and Brownian Ratchets

Week 5: The rate equation paradigm and genetic networks

Week 6: Noise in biological systems

Week 7: Turing patterns in embryogenesis

Week 8: Mechanics in embryogenesis and Future directions

## **Books and references**

- 1. Howard Berg (1993) Random Walks in Biology Princeton Univ. Press. ISBN 9780691000640
- Philip Nelson (2007) Biological Physics: Energy, Information, Life. W. H. Freeman. ISBN-13: 978-0716798972
- 3. Rob Philips, Jane Kondev, JulieTheriot, Hernan Garcia (2013) Physical Biology of the cell. CRC Press. ISBN 9780815344506
- 4. David Boal (2012) Mechanics of the Cell. 2nd edition, Cambridge University Press. Online ISBN: 9781139022217

 Gabor Forgacs and Stuart Newman (2005) Biological Physics of the Developing Embryo. Cambridge University Press. Online ISBN: 9780511755576